



REVIEW

Structure-function relationship of the mammarenavirus envelope glycoprotein

Wei Wang[✉], Zheng Zhou, Leike Zhang, Shaobo Wang, Gengfu Xiao

State Key Laboratory of Virology, Wuhan Institute of Virology, Chinese Academy of Sciences, Wuhan 430071, China

Mammarenaviruses, including lethal pathogens such as Lassa virus and Junín virus, can cause severe hemorrhagic fever in humans. Entry is a key step for virus infection, which starts with binding of the envelope glycoprotein (GP) to receptors on target cells and subsequent fusion of the virus with target cell membranes. The GP precursor is synthesized as a polypeptide, and maturation occurs by two cleavage events, yielding a tripartite GP complex (GPC) formed by a stable signal peptide (SSP), GP1 and GP2. The unique retained SSP interacts with GP2 and plays essential roles in virion maturation and infectivity. GP1 is responsible for binding to the cell receptor, and GP2 is a class I fusion protein. The native structure of the tripartite GPC is unknown. GPC is critical for the receptor binding, membrane fusion and neutralization antibody recognition. Elucidating the molecular mechanisms underlining the structure–function relationship of the three subunits is the key for understanding their function and can facilitate novel avenues for combating virus infections. This review summarizes the basic aspects and recent research of the structure–function relationship of the three subunits. We discuss the structural basis of the receptor-binding domain in GP1, the interaction between SSP and GP2 and its role in virion maturation and membrane fusion, as well as the mechanism by which glycosylation stabilizes the GPC structure and facilitates immune evasion. Understanding the molecular mechanisms involved in these aspects will contribute to the development of novel vaccines and treatment strategies against mammarenaviruses infection.

KEYWORDS Mammarenaviruses; glycoprotein complex (GPC); stable signal peptide (SSP); membrane fusion; glycosylation

INTRODUCTION

Mammarenaviruses belong to the genus *Mammarenavirus*, family *Arenaviridae*, which consists of 27 unique species currently recognized by the International Committee on Taxonomy of Viruses (ICTV) (Radoshitzky et al., 2015). The original classification of mammarenaviruses, based mainly on virus antigenic properties, serological,

genetic and geographical relationships, and the rodent host, identified two groups: New World (NW; also referred to as Tacaribe serocomplex) and Old World (OW; also referred to as Lassa-lymphocytic choriomeningitis serocomplex) mammarenaviruses (Salvato et al., 2005). The NW mammarenavirus are further divided into clades A, B, C and D, in which clade D was once named as a recombinant A/B (recA) clade (Bowen et al., 1997; Clegg, 2002; Emonet et al., 2006). The phylogenetic tree of mammarenavirus recognized by the ICTV, as well as some novel arenaviruses, is shown in [Figure 1](#). The family prototype, lymphocytic choriomeningitis virus (LCMV), was first isolated in 1933 during serial passages in monkeys of human material obtained from a fatal infection in the first documented epidemic of St. Louis enceph-

Received: 20 May 2016, Accepted: 27 June 2016,
Published online: 4 August 2016

✉Correspondence:
Phone: +86-27-87198352, Fax: +86-27-87198685,
Email: wangwei@wh.iov.cn
ORCID: 0000-0002-8696-9773

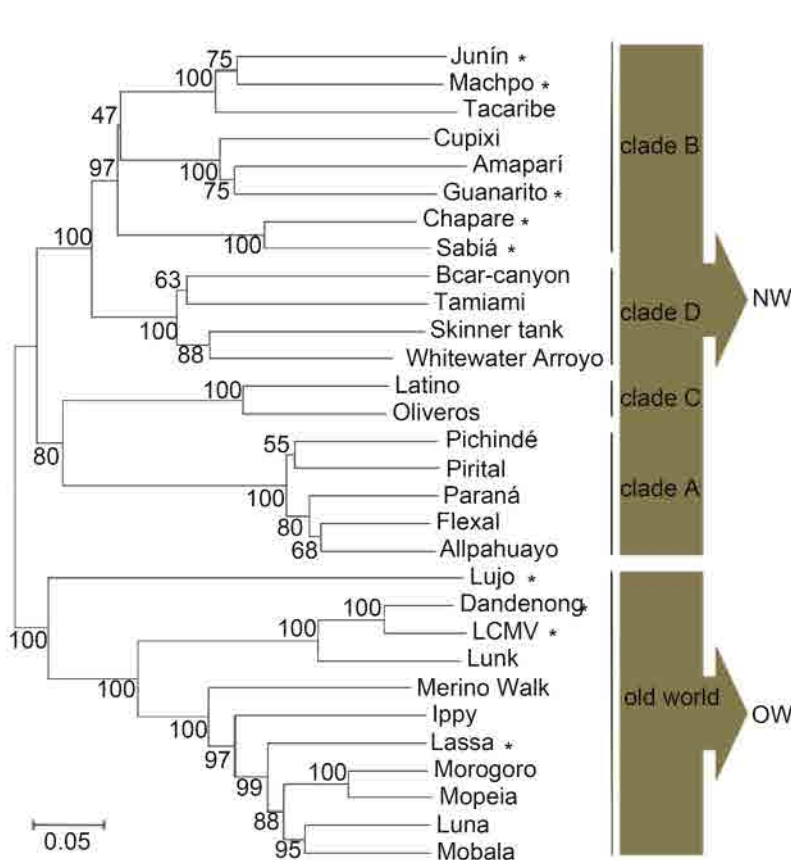


Figure 1. Mammarenavirus phylogenetic tree. Amino acid sequences of mammarenavirus GPCs were aligned using ClustalX. The phylogenetic tree was computed using the pairwise distance algorithm with neighborjoining analysis in MEGA 4.0 software. Support for nodes was confirmed by bootstrapping using 500 pseudoreplications. The UniProt accession codes included are Q90037 (Sabiá), Q84168 (Oliveros), Q911P0 (Whitewater Arroyo), Q27YE4 (Ippy), Q9DK03 (Allpahuayo), Q8B122 (Amaparí), Q8B121 (Latino), Q8B120 (Paraná), Q8B119 (Pirital), Q8B115 (Cupixi), Q8AZ52 (Flexal), Q8AYY5 (Tamiami), Q8AYW1 (Guanarito), Q6IUF7 (Machupo), Q2A069 (Mobala), P26313 (Junín), P19240 (Mopeia), P18141 (Tacaribe), P08669 (Lassa), P07399 (LCMV, lymphocytic choriomeningitis virus), P03540 (Pichindé), G3XE6 (Luna), D4N7Z6 (Merino Walk), C6ZK00 (Morogoro), C5ILC1 (Lujó), B2KSC1 (Skinner tank), B2C4J0 (Chapare), B1NX58 (Dandenong), A0PJ25 (Bearcanyon) and K0IT71 (Lunk). Skinner tank virus, Morogoro virus, and Dandenong virus have not yet been recognized by the International Committee on Taxonomy of Viruses. Pathogenic mammarenavirus are indicated by an asterisk.

alitis (Armstrong and Lillie, 1934; Oldstone, 2007). LCMV is the only mammarenavirus to exhibit a worldwide distribution due to its association with the common house mouse, *Mus musculus*. Other mammarenaviruses are distributed in North America, South America or Africa. The OW and clade B groups contain the major viruses causing hemorrhagic fevers (HF) on the African continent and in South America, respectively, as well as non-pathogenic viruses. Lassa virus (LASV), Junín virus (JUNV), Machupo virus (MACV), Guanarito virus (GTOV) and Sabiá virus (SABV) are known to cause severe hemorrhagic fever in western Africa, Argentina, Bolivia, Venezuela and Brazil, respectively (Buchmeier et al., 2007). They are included in the Category A Pathogen List as defined by the Centers for Disease Control and Prevention, and are listed as Biosafety Level 4 (BSL-4) agents.

Mammarenavirus virions are enveloped viruses containing a bisegmented, ambisense RNA genome with four open reading frames. The long (L) RNA segment ($\approx 7,200$ nt) encodes the viral RNA-dependent RNA polymerase (L protein, 200 kDa) and a small RING-finger matrix protein (Z protein, 11 kDa). The short (S) segment ($\approx 3,500$ nt) encodes the nucleoprotein (NP, 63 kDa) and the viral glycoprotein complex (GPC) precursor (75 kDa). The L protein is responsible for transcribing viral mRNAs and rep-

licating the genome (Leung et al., 1979; Fuller-Pace and Southern, 1989; Garcin and Kolakofsky, 1990; Kranzusch and Whelan, 2012). The matrix protein Z plays key roles in budding (Salvato and Shimomaye, 1989; Salvato et al., 1992; Perez et al., 2003; Strecker et al., 2003; Eichler et al., 2004b). The structural NP encapsidates the genome segments and is essential for transcription and replication (Pinschewer et al., 2003).

The GPC precursor is expressed as a polyprotein and matures by undergoing two cleavage events. First, the GPC precursor is cleaved by cellular signal peptidases (SPases) in the endoplasmic reticulum (ER) (Eichler et al., 2003a, b), where the glycoprotein (GP) is extensively *N*-glycosylated. Unlike most other viruses, the cleaved 58 aa stable signal peptide (SSP) is not degraded but is retained with the immature GP precursor (York and Nunberg, 2007a). The second cleavage event takes place later in the Golgi/trans-Golgi network; the exception is LASV, whose GP is cleaved early in the ER/*cis*-Golgi compartment (Lenz et al., 2001; Burri et al., 2012; da palma et al., 2014). The cellular proprotein convertase subtilisin kexin isozyme-1 (SKI-1)/site-1 protease (S1P) enzyme recognizes the motif RX(hydrophobic)X ↓ and cleaves the immature GP1/2 into the GP1 and GP2 subunits (Lenz et al.,

2001; Beyer et al., 2003; Kunz et al., 2003; Rojek et al., 2008a). In this motif, X is any amino acid except cysteine and the hydrophobic amino acid is preferentially leucine. In OW viruses, the cleavage motif is “RRLX ↓” except for Lujo virus (Pasquato et al., 2011; Burri et al., 2013). The motif present in the LASV GPC is RRLI, which has been

proved to be crucial for the cleavage in the ER (Burri et al., 2012). The three non-covalently bound subunits-SSP, GP1 and GP2-form a (SSP/GP1/GP2)₃ trimeric complex and traffic to the cell surface for virion budding and release (Young and Howard, 1983; Burns and Buchmeier, 1991; Eschli et al., 2006) (Figure 2).

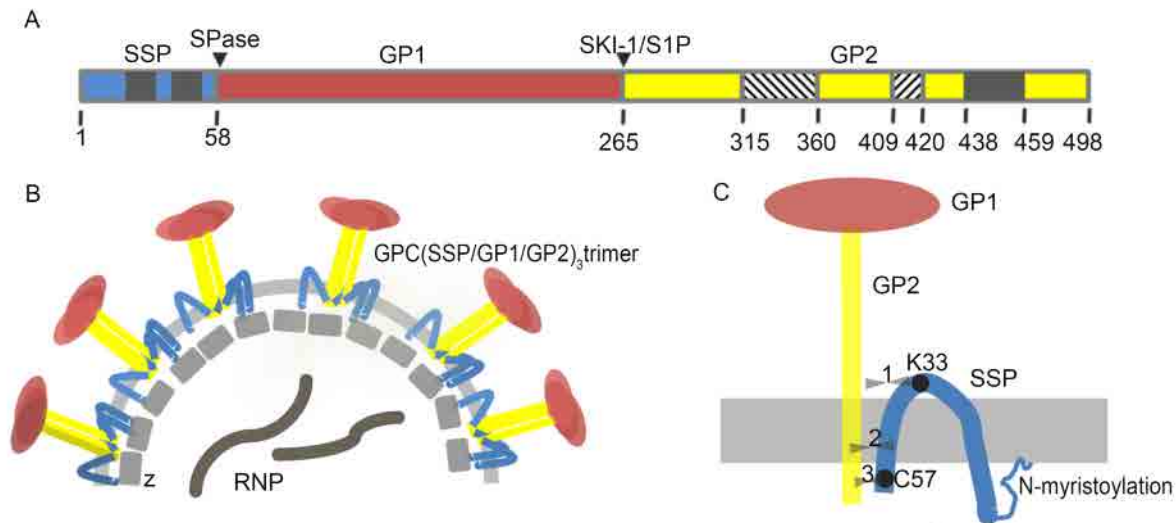


Figure 2. Schematic diagrams of the mammarenavirus envelope GPC. (A) Schematic diagram of the LCMV GPC open reading frame, with the cleavage sites of SPase and SKI-1/S1P indicated. The transmembrane domains in SSP and GP2 are shown in gray. The N- and C-terminal helices in GP2 are shown as lower diagonal and upper diagonal hatching, respectively. (B) Virion structure showing the tripartite GPC, Z protein and ribonucleoprotein (RNP) complex containing NP, L protein and the viral genome. (C) Detailed schematic diagram of the SSP/GP1/GP2 heterotrimer. The three opposing arrowheads represent the potential interaction sites between SSP and GP2, which are discussed in detail in the text. The drawing is not to scale.

The unique SSP/GP1/GP2 heterotrimer of mammarenavirus GPCs is very different from other viruses. The retained SSP plays essential roles in GPC biosynthesis, cleavage and maturation, as well as fusion activity (Eichler et al., 2003a; York and Nunberg, 2006; Agnihothram et al., 2007; Saunders et al., 2007; York and Nunberg, 2007a; Messina et al., 2012; Bederka et al., 2014). GP1 is the distal protein on the virion spike and is responsible for receptor recognition and binding (Cao et al., 1998; Abraham et al., 2010). GP2 mediates membrane fusion after the virions are internalized into acidic endosomes (York et al., 2005; Harrison, 2008; Rojek and Kunz, 2008). According to the sequence alignment results, SSP and GP2 are highly conserved, with amino acid identities of 67.3% and 70.51%, respectively (calculated by DNAMAN software). The alignment of GP1 reveals it is less conserved and the amino acid identity is about 33%. The interactions among the three subunits of the GPC are complex. Recently, the crystal structure of the LCMV GP1/GP2 prefusion complex was reported (Hastie et al., 2016). The extending GP1 N terminus interacts with the C helix, two β -strands and fusion loop from GP2, to form the important conforma-

tional epitope that is recognized by most neutralizing antibodies (nAbs) (Figure 3) (Robinson et al., 2016). The interaction of GP1 with the receptor initiates endocytosis. In the endocytic pathway, the acidic pH induces the fusogenic conformational change of GP2 and exposure of the fusion region. Then the fusion region inserts into the cell membrane and GP2 undergoes conformational rearrangements, which is the driving force for membrane fusion (Borrow and Oldstone, 1994). Meanwhile, the interaction between SSP and GP2 is important for pH-induced membrane fusion (York and Nunberg, 2009), but the mechanism is not yet fully understood. Elucidating the molecular mechanisms underlining the structure-function relationship of the three subunits is the key for understanding their function and may facilitate novel avenues for combating virus infections.

GP1 IS RESPONSIBLE FOR THE RECEPTOR BINDING

Mammarenavirus infection is initiated by the binding of

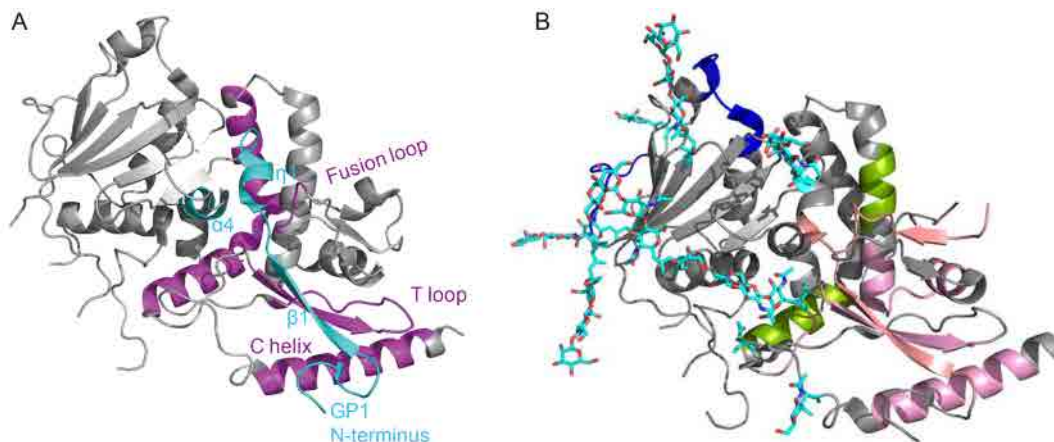
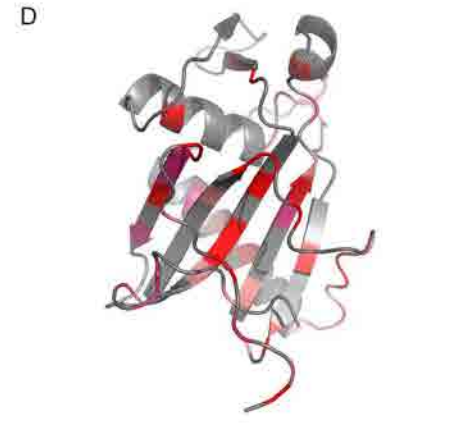
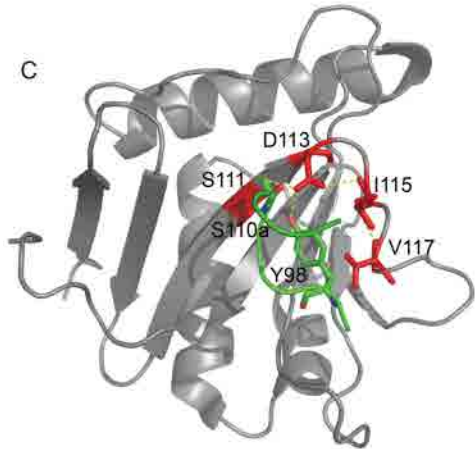
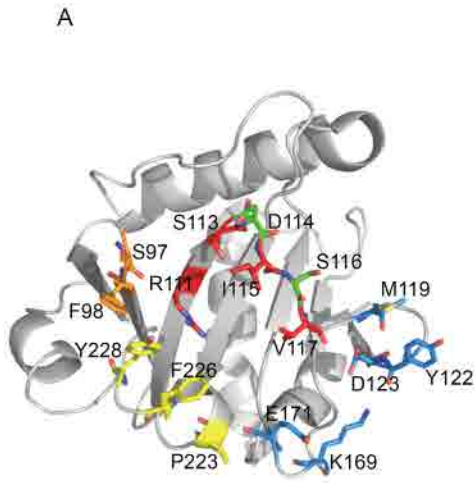


Figure 3. Crystal structure of the LCMV GP1 and GP2 prefusion complex. The Protein Data Bank (PDB) accession number is 5INE. (A) Cartoon diagram of LCMV GP1–GP2 interactions. The extending GP1 N terminus, N-terminal loop, $\beta 1$ and $\eta 1$ (cyan) interact with the C-terminal helix, two β -strands and the fusion loop from GP2 (purple), respectively. $\alpha 4$ in GP1 (cyan) occupies a cleft (purple) in GP2. (B) Putative epitopes are indicated by light red, green, blue and pink highlights. N-Linked glycans visible in the GP1–GP2 crystal structure are illustrated as cyan ball-and-stick models with the linked asparagine.

GP1 to target cell surface receptors, followed by internalization by endocytosis (Borrow and Oldstone, 1994; Martinez et al., 2007; Vela et al., 2007) and pH-dependent fusion with late endosomes, with the subsequent release of the virus ribonucleoprotein complex into the cytoplasm (Meyer et al., 2002). The first identified receptor for arenavirus was α -dystroglycan (α -DG), a highly glycosylated membrane protein acting as a ubiquitous receptor (Cao et al., 1998). α -DG has been identified as the receptor for both OW and clade C mammarenavirus (Rojek et al., 2007a, b), and *O*-mannosylation modification of DG was reported to be necessary for LASV binding (Kunz et al., 2005; Hara et al., 2011; Jae et al., 2013). It was shown recently that successful infection of LASV requires a receptor switch from α -DG to a lysosomal protein termed LAMP1 in a pH-dependent manner (Jae et al., 2014). Among the NW clade B mammarenavirus, pathogenic viruses use human transferrin receptor 1 (hTfR1) for entry (Radoshitzky et al., 2007; Flanagan et al., 2008). Non-pathogenic clade B viruses use TfR1 orthologs but are unable to gain cellular entry via hTfR1 (Abraham et al., 2009). However, infection by Lujo pseudotype virus uses neither hTfR1 nor α -DG (Tani et al., 2014). As shown in Figure 4E, Lujo virus GP1 has a low amino acid homology with other OW mammarenaviruses, which might make the receptor-binding sites significantly different from others. Recently, some novel receptor candidates for JUNV, LASV and LCMV have been identified, such as C-type lectin family, dendritic cell-specific intercellular adhesion molecule 3-grabbing nonintegrin (DC-SIGN), liver and lymph node sinusoidal endothelial calcium-dependent lectin (LSECtin), and two members of the TAM family (Axl and Tyro3)

(Shimajima and Kawaoka, 2012; Shimajima et al., 2012; Goncalves et al., 2013; Martinez et al., 2013). Although α -DG and TfR1 are the major receptors, mammarenaviruses may enter into cells in several ways using different molecules, especially in those cells showing low expression of α -DG or TfR1 (Bedossa et al., 2002; Yamamoto et al., 2004).

The crystallographic structure of MACV GP1 was solved for the unbound protein (Bowden et al., 2009) and for its complex with TfR1 (Abraham et al., 2010). The Protein Data Bank (PDB) code for MACV GP1 is 2WFO (GP1 aa 87–239). MACV GP1 is monomeric when crystallized, with a novel α/β -fold, and is stabilized by four disulfide bonds around the overall folds (Figure 4A, 4B). The relatively conserved disulfide bond presented in MACV, JUNV and Tacaribe virus (TCRV) GP1 reflects the close relationship of these three viruses, as shown in Figure 1 (Bowen et al., 1996; Charrel et al., 2002; Cajimat et al., 2009). The co-crystallization structure of MACV GP1 with hTfR1 has also been depicted (PDB code 3KAS). MACV GP1 interacts with the tip of the apical domain of hTfR1. Five interaction motifs in the interface have been identified, as shown in Figure 4A, 4B (Abraham et al., 2010). Some residues in the motifs, such as R111, Y122 and F226, are essential for GP1-TfR1 interaction, while some residues, such as D114 and S116, are expendable. Besides these five motifs, some residues, such as D155 and P160, have been shown to be important for the GP1-TfR1 interaction in NW clade B mammarenaviruses (Radoshitzky et al., 2011). They may aid in the stabilization of GP1 folding and the GP1-TfR1 interaction. Because clade B contains the major pathogenic mammar-



B

	60	70	80	90	β1	100	β2	TT	β3		
MACV	CSDG	EIKIGLHTEFC	SVTLTMQR	L	ANHSN	ELP	SLC	MLNS	NSFY	MRGGVNT	...FL
GTOV	...	IKVCHHTNF	SFTVLI	GGVF	...	HEL	PL	CL	CRVNS	YSYI	IRLSHNS
CPKV	...	RIKCHHTNF	SFTMTI	GGVF	...	HEL	PL	CL	CRVNS	YSYI	IRLSHNS
AMAV	...	RIKCHHTNF	SFTMSV	GGVF	...	HEL	PL	CL	CRVNS	YSYI	IRLSHNS
CHPV	...	RSCS	EIKIGR	TELO	NI	TINML	KVF	...	ED	HP	ISCT
SABV	...	RIKGR	TELO	NI	TIDML	KVF	...	ED	HP	ISCT	MYNH
TACV	...	EET	EIKIGH	TEK	QV	SLSL	SALL	...	IN	QSH	ELP
JUNV	...	EEAD	EIKIGH	TEK	QV	SSSM	VGL	...	SN	NP	DL

	110	120	β4	η1	η2	α1
MACV	TRVSD	IVLMK	...	EYD	V	SIYEPED
GTOV	WEYVD	HPVLCSS	...	S	F	HLIDNY
CPKV	WEYVD	HPVLCSS	...	S	F	HLIDNY
AMAV	WEYVD	HPVLCSS	...	S	F	HLIDNY
CHPV	WEYVD	HPVLCSS	...	S	F	HLIDNY
SABV	WEYVD	HPVLCSS	...	S	F	HLIDNY
TACV	WEYVD	HPVLCSS	...	S	F	HLIDNY
JUNV	WEYVD	HPVLCSS	...	S	F	HLIDNY

	TTT	β5	TT	TT	*	180	α2
MACV	CHDW	YLD	DP	PP	ML	CRN	K
GTOV	CHDF	YLD	DP	PP	ML	CRN	K
CPKV	CHDF	YLD	DP	PP	ML	CRN	K
AMAV	CHGL	YLD	DP	PP	ML	CRN	K
CHPV	KYDF	YLD	DP	PP	ML	CRN	K
SABV	KYDF	YLD	DP	PP	ML	CRN	K
TACV	KYDF	YLD	DP	PP	ML	CRN	K
JUNV	CHDW	YLD	DP	PP	ML	CRN	K

	210	β7	TT	η3	α3	230	240	250	260
MACV	EEFG	...	KVCY	IT	IN	Q	GGP	SS	F
GTOV	SYIS	...	KMS	V	MR	NT	T	W	E
CPKV	KDP	...	KIF	V	MR	NT	T	W	E
AMAV	KEG	...	KD	V	IL	R	N	T	T
CHPV	KTP	...	KQ	F	V	I	R	N	T
SABV	NIQD	...	L	G	K	R	F	L	L
TACV	FDG	...	E	Q	C	K	I	Q	E
JUNV	LDG	...	K	I	C	L	M	K	A

E

	60	70	80	TT	β1	100	β2	110
LASV	TS	LYK	GVY	EL	Q	T	L	E
Dandenong	PD	IY	K	G	V	Y	F	L
LCMV	PD	IY	K	G	V	Y	F	L
Morogoro
MOPV
MBLV
Luna
Ippy
Merino
Lujo

	120	α1	130	140	150	β3	TT	β4	170
LASV	FC	N	L	S	D	A	H	K	
Dandenong	FC	N	L	S	D	A	H	K	
LCMV	FC	N	L	S	D	A	H	K	
Morogoro	FC	N	L	S	D	A	H	K	
MOPV	FC	N	L	S	D	A	H	K	
MBLV	FC	N	L	S	D	A	H	K	
Luna	FC	N	L	S	D	A	H	K	
Ippy	FC	N	L	S	D	A	H	K	
Merino	FC	N	L	S	D	A	H	K	
Lujo	FC	N	L	S	D	A	H	K	

	α2	180	α3	190	200	β5	T	T	β6	β7
LASV	DA	AN	H	C	G	T	V	A		
Dandenong	DA	AN	H	C	G	T	V	A		
LCMV	DA	AN	H	C	G	T	V	A		
Morogoro	DA	AN	H	C	G	T	V	A		
MOPV	DA	AN	H	C	G	T	V	A		
MBLV	DA	AN	H	C	G	T	V	A		
Luna	DA	AN	H	C	G	T	V	A		
Ippy	DA	AN	H	C	G	T	V	A		
Merino	DA	AN	H	C	G	T	V	A		
Lujo	DA	AN	H	C	G	T	V	A		

	230	240	250	260
LASV	WE	D	H	C
Dandenong	WE	D	H	C
LCMV	WE	D	H	C
Morogoro	WE	D	H	C
MOPV	WE	D	H	C
MBLV	WE	D	H	C
Luna	WE	D	H	C
Ippy	WE	D	H	C
Merino	WE	D	H	C
Lujo	WE	D	H	C

Figure 4. Structure of MACV, JUNV and LASV GP1. (A) The five receptor interaction motifs are shown in red, green, yellow, blue and orange, respectively. (B) Sequence alignment of MACV GP1 aa 77–262 with other NW clade B mammarenaviruses: Chapare virus (CHPV), SABV, Cupixi virus (CPXV), Amapari virus (AMAV), GTOV, JUNV and Tacaribe virus (TCRV). The observed secondary structure is indicated above the sequence. Disulfide bonds are numbered below the alignment. Residues of the five interaction motifs are colored as in (A). Secondary structure elements are graphically represented by ESPript (Robert and Gouet, 2014). (<http://esprript.ibcp.fr>) (C) Cartoon diagram of JUNV GP1 with the complementarity-determining region of nAb GD01 (residues 97–100 aa) shown as sticks. (PDB code 5EN2). (D) Cartoon diagram of LASV GP1 (PDB code 4ZJF). LASV GP1 is colored by a relative conservation score based on multiple-sequence alignment of OW mammarenavirus GP1 sequences, which was calculated using the ConSurf server (Landau et al., 2005). (E) Sequence alignment of LASV GP1 with other OW mammarenaviruses: LCMV, Dandenong virus, Mopeia virus (MOPV), Morogoro virus, Mobala virus (MBLV), Luna virus, Ippy virus, Merino walk virus and Lujo virus, showing the secondary structure elements as observed in the crystal structure of LASV GP1. Fully conserved residues are highlighted with a red background, and partially conserved residues are shown in red.

enaviruses, these conserved and critical residues might provide a novel perspective for developing broad-spectrum inhibitors against HF.

Identification of the receptor-binding domains (RBDs) is critical for drug development and antibody design (Lalezari et al., 2003; Lazzarin et al., 2003; Dorr et al., 2005; Bossart et al., 2009; Zu et al., 2014). nAb targeting the RBD of filovirus blocks both Marburg virus and Ebola virus entry (Flyak et al., 2015; Hashiguchi et al., 2015), suggesting that RBD is a candidate target for broad-spectrum inhibitor design. The crystal structure of JUNV GP1 bound to the nAb has been reported (PDB code 5EN2; Mahmutovic et al., 2015). The molecular mechanism of antibody-mediated neutralization revealed that the antibody occupied the RBD of JUNV GP1, which was prevented from binding to the cell receptor (Figure 4C) (Mahmutovic et al., 2015).

The crystal structure of LASV GP1 in an acidic environment has been reported (PDB code 4ZJF, GP1 aa 75–237; Cohen-Dvashi et al., 2015). Structure alignment of LASV GP1 and MACV GP1 has revealed that the most structurally conserved region is the central part of the core β -sheet, while highly diversified structures exist at the termini of LASV GP1, which poses a steric barrier that precludes the binding sites contacting TRf1 (Cohen-Dvashi et al., 2015) (Figure 4D). This may reflect the evolutionary distance between OW and NW viruses. However, the reported structure of LASV GP1 obtained at acidic pH could pull down endogenous LAMP1 at pH 5.0 but not α -DG at pH 5.0 or at pH 8.0, indicating that the crystal structure is different from the native structure on the virion spike, which confers a LAMP1-compatible state (Cohen-Dvashi et al., 2015). Although a putative LAMP1-binding motif centered by three histidines (H92, H93 and H230) is conserved in OW viruses (Cohen-Dvashi et al., 2015) (Figure 4E), LCMV has been shown not to require LAMP1 for infection of cells (Jae et al., 2014). The structure of an intact trimeric LASV GPC complex was observed recently by low-resolution electron cryomicroscopy. The tripartite

GPC forms the surface spike, which is distributed irregularly on the virion envelope and is connected to the matrix. Following virus internalization, the acidic pH causes GP1 to undergo conformational changes exposing the histidine triad for LAMP1 binding (Li et al., 2016).

The RBD of GP1 bound to α -DG has yet been reported. It has been demonstrated that *O*-glycan biosynthesis factors, such as the transferase like-acetylglucosaminyl-transferase (LARGE), which glycosylates α -DG with *O*-mannosyl glycans, are indispensable for virus binding (Imperiali et al., 2005; Kunz et al., 2005). Moreover, the interaction of LASV GP1 with LAMP1 is also dependent on the glycans linked to LAMP1 (Jae et al., 2014). Most recently, crystallization of the prefusion LCMV GP1-GP2 complex revealed five critical receptor-binding residues located in or near the basic helix-loop face, which might interact with the acidic sugar on the receptor surface (Hastie et al., 2016). Interesting, in NW viruses such as MACV, GP1 is necessary and sufficient for receptor binding, while in OW viruses such as LASV or LCMV, only the full-length GPC, and not GP1 alone, can achieve α -DG engagement (Jae et al., 2014; Cohen-Dvashi et al., 2015; Hastie et al., 2016). This is similar to the human nAb against LASV. Most human nAbs recognize the conformational epitope consisted of the extended N terminus of GP1 and parts of GP2 that interact with the N terminus of GP1 (Figure 3A) (Robinson et al., 2016).

GP2 IS THE MEMBRANE FUSION PROTEIN

GP2, the membrane fusion subunit of the GPC, belongs to the class I viral fusion proteins, presenting a typical α -helix-rich trimer in its postfusion state (Eichler et al., 2006; Klewitz et al., 2007). In the recently reported prefusion structure of LCMV GP2, several striking differences were observed compared with the postfusion structure. In the prefusion state, the long-span N-terminal helix in the postfusion structure was seen as four segments, the T-loop formed antiparallel β -strands instead of the two α -helices

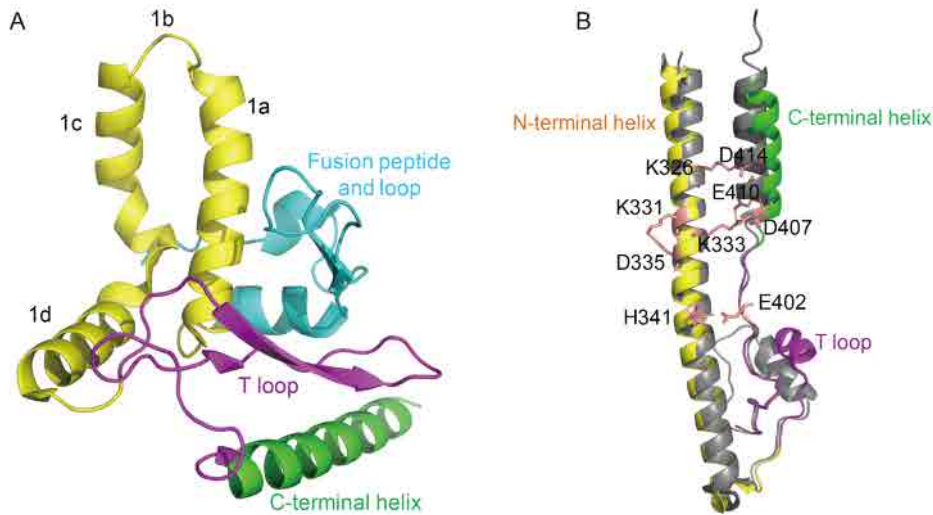


Figure 5. Structure of mammarenavirus GP2. (A) Cartoon diagram of LCMV GP2 in the prefusion state (PDB code 5INE). (B) Superposition of postfusion GTOV GP2 (gray, PDB code 4C53) with LCMV GP2 (colored as in (A), PDB code 3MKO), generated by the PyMOL Molecular Graphics System, version 1.2r3pre (Schrödinger, LLC). Residues involved in intra-chain salt bridges are shown as salmon ball-and-sticks.

in the postfusion structure, and the C-terminal helix formed hydrophobic contacts with the extending N terminus of GP1 (Figure 5A) (Igonet et al., 2011; Hastie et al., 2016). During entry, the gradual acidification triggers a fusogenic conformational change of GP2 (Disimone et al., 1994). The C-terminal helix and N-terminal helix form a postfusion hairpin, which is stabilized by intra-chain salt bridges, and three hairpins gather and form the typical six-helix bundle of a class I fusion protein (Figure 5B). The fusion region of LCMV GP2 has a bipartite region that includes an N-terminal fusion peptide and an internal fusion loop, which is thought to be identical across mammarenaviruses.

In the postfusion state, the inter-chain salt bridge, formed by D353 and R358, and the strictly conserved N325, chelating the central chloride ion, stabilizes the postfusion LCMV GP2 coiled-coil trimer (Igonet et al., 2011). Based on the high amino acid sequence homology, GP2 of OW viruses is suggested to form a conserved structure. Crystallization of the postfusion construct of GTOV GP2 (PDB code 4C53), which belongs to the NW viruses, revealed that it is identical in length to LCMV GP2, with both omitting the fusion region to avoid aggregation, and that they have about 50% amino acid sequence identity and exhibit very similar secondary structures (Parsy et al., 2013) (Figure 5B). Unlike other class I viral fusion proteins, GP2 of some pathogenic mammarenaviruses such as LASV and JUNV mediates membrane fusion at a remarkably low pH optimum (< 4.5 for LASV and < 5.5 for JUNV) (York and Nunberg, 2006; Klewitz et al., 2007; Quirin et al., 2008; Rojek et al., 2008b; Cosset et al., 2009; Pasqual et al., 2011). Although the GP2 sequences show high amino acid sequence homology, some

minor structural differences might attribute to the difference in sensitivity to acidic pH in late endosomes or lysosomes and the subsequent membrane fusion.

SSP IS UNIQUE IN MAMMARENAVIRUS ENVELOP GLYCOPROTEIN

Mammarenavirus SSP is essential for virion maturation and subsequent infection (Bederka et al., 2014). As shown in Figure 6, SSP exhibits high amino acid sequence conservation in mammarenaviruses. The 58 aa SSP contains two hydrophobic transmembrane domains (TM1, aa 18–32; and TM2, aa 41–54) linked by a short ectodomain (Figure 2) (Eichler et al., 2003b; Froeschke et al., 2003; Eichler et al., 2004a; Agnihothram et al., 2007; Saunders et al., 2007; Schrepf et al., 2007). SSP plays essential roles in GPC cleavage maturation. SSP association is required in JUNV to allow the exit of the pre-maturation GPC (pGPC) from the ER and thus for exposure to the SKI-1/S1P protease in the Golgi (Agnihothram et al., 2006). The myristoylated N terminus is assumed to be anchored in the intracellular membrane (York et al., 2004). G2A mutation in the myristoylated motif of LCMV SSP causes reduced virus-like particle (VLP) fusion as well as entry, and blocks GPC-mediated fusion but retains all other SSP functions (Saunders et al., 2007). Similarly, lack of SSP myristoylation in JUNV prevents membrane fusion, with no effect on the formation of the (SSP/GP1/GP2)₃ trimer (York et al., 2008). The penultimate C-terminal C57 is associated with the zinc-finger motif in GP2, which is essential for stabilizing SSP incorporation in GPC (York and Nunberg, 2007b; York and Nunberg,

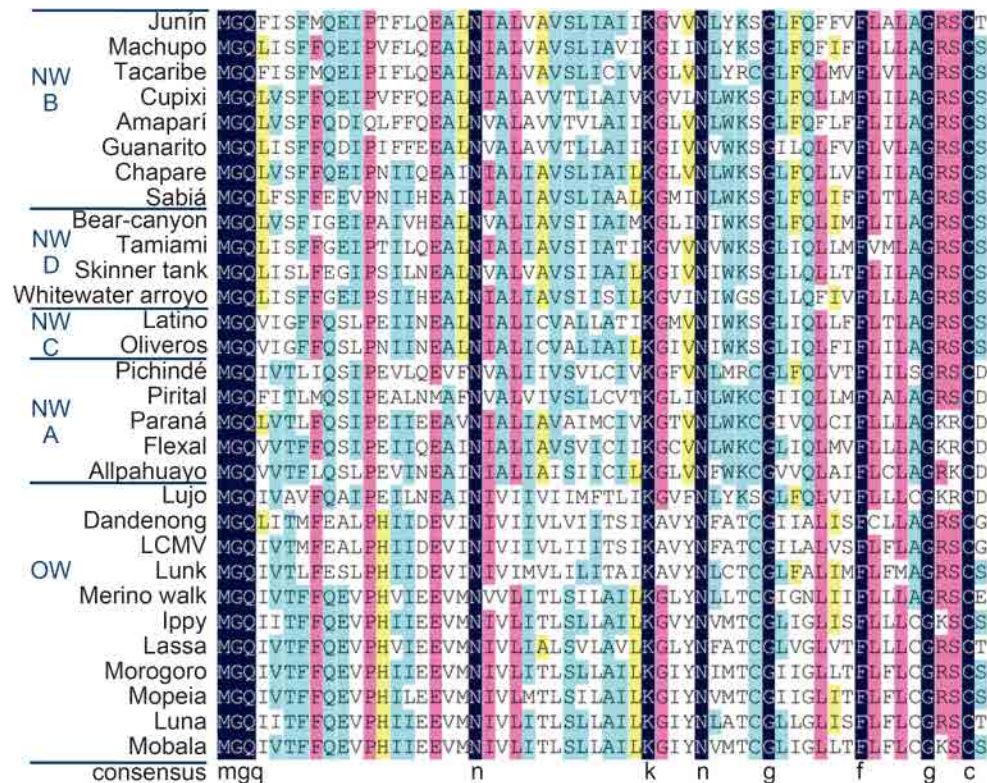


Figure 6. Alignment of mammarenavirus SSP sequences. Sequence alignment was analyzed using DNAMAN, with 100% amino acid identity shown in dark blue, ≥ 75 in pink, $\geq 50\%$ in blue and $\geq 33\%$ in yellow. Strictly conserved residues such as the myristoylation motif, K33 and C57 are indicated below the sequences.

2007a; Briknarova et al., 2011).

Homotypic pairing of the SSP N-terminal cytoplasmic region with the TM1 region (aa 1–32) is paramount for membrane fusion activity. However, homology in the ectodomain, TM2 and the C-terminal cytoplasmic regions appears to be relatively unimportant in fusion activity (Messina et al., 2012). In LASV, pGPC with deletion of either SSP TM1 or TM2 can still traffic to the ER, but deletion of TM2 leads to a reduction of SSP cleavage (Eichler et al., 2004a), which is a prerequisite for GPC maturation (Eichler et al., 2003a). However, somewhat larger TM1 and TM2 deletion mutations (aa 10–32 and aa 34–53, respectively) eliminates virtually all SSP functions.

A conserved hydrophobic motif (FLLL) upstream of the SPase cleavage site is assumed to be the key for GP maturation and trafficking. Mutation of this motif leads to the impairment of GP maturation processing, a reduction of surface expression and the impedance of membrane fusion. Moreover, each of these four residues in this motif has been shown to play an equal role in GPC maturation cleavage and expression. The phenylalanine at position 49 in this motif has also been confirmed to affect both acidic fusion and infectivity (Agnihothram et al., 2007; Saunders et al., 2007).

Some strictly conserved residues across the mammarenaviruses are not only indispensable for SPase cleavage but are critical for SSP function (York and Nunberg, 2007a). A conserved proline at position 12 is essential for infectivity. Both P12A and P12G mutants disrupt VLP fusion and entry. However, a P12G mutant increases membrane fusion activity, while a P12A mutant decreases this, suggesting that SSP plays different roles in GPC-mediated fusion activity and infectivity (Saunders et al., 2007). Moreover, P12 is suggested to be involved in a viral budding motif, Φ PXV, which is critical for paramyxovirus budding. In this motif, Φ is any aromatic amino acid and X is any amino acid (Schmitt et al., 2005). The sequence conserved across mammarenaviruses is (I/L/V)PX(hydrophobic residue).

The invariably conserved asparagine residues located at 20 and 37 in SSP have been shown to be essential for GPC function. N20 is critical for VLP fusion and entry, while both N20 and N37 are required for GPC maturation (York and Nunberg, 2006). In LCMV, a K33R mutation retains a wild-type level of pGPC expression and cell surface transport, as well as GPC cleavage and maturation. Positive-charge mutations at position 33 are better than no-charge mutations, and no-charge mutations are still

better than the negative-charge mutations for GPC maturation and downstream function. This positive-charge preference at position 33 has also been demonstrated in JUNV SSP (York and Nunberg, 2006). Moreover, K33 in the SSP ectodomain is crucial for GPC sensitivity to acidic pH and subsequent membrane fusion, as discussed above. As indicated above, a K33Q mutant is deficient in membrane fusion. However, mutations in the membrane-proximal external region (MPER) (D400A, E410A, R414A and K417A) in JUNV GP2 were shown to restore this deficiency of pH sensitivity (York and Nunberg, 2009). Hydrophobic residues in the transmembrane domain (F427, W428 and F438) in GP2 also exhibit such complementation, and might interact with residues in the SSP transmembrane domain to stabilize GPC. Interestingly, JUNV K33R, K33H, K33Q and K33E mutations acquire resistance to ST-294, a small-molecule entry inhibitor of NW mammarenaviruses preventing membrane fusion by targeting the SSP–GP2 interface (York et al., 2008).

THE SSP-GP2 INTERACTION STABILIZES GPC AND PROVIDES A TARGET FOR DRUG DESIGN

Although the sequence identity of SSP and GP2 in mammarenaviruses is high, hybrid GPC complexes (JUNV SSP with LASV GP1/GP2, or LASV SSP with JUNV GP1/GP2) are unable to induce membrane fusion (Albarino et al., 2011b). However, the hybrid GPC is still correctly assembled and maturation still occurs by cleavage with SPase and SKI-1/S1P (Albarino et al., 2011b; Messina et al., 2012). The mechanism of membrane fusion is not only associated with the conformational changes of GP2 but is also related to the unique SSP subunit, which is much different from other class I fusion proteins. The specific SSP and GP2 interaction is essential for stabilizing the prefusion GPC complex. It is reasonable to believe that SSP interacts with GP2 at ectodomain, transmembrane domain and C-terminal cytoplasmic domain (Figure 2C).

Candid #1, a live-attenuated JUNV vaccine, is currently the only vaccine available against an arenavirus and is licensed for human use in Argentina (Maiztegui et al., 1998; Enria and Barrera Oro, 2002). Candid #1 has the major attenuating mutation F427I in the transmembrane domain of GP2. This critical residue is suggested to incorporate with SSP to sustain the high-energy GPC trimeric conformation (Albarino et al., 2011a; Droniou-Bonzom et al., 2011; Seregin et al., 2015). Furthermore, York and Nunberg (2007b) identified six conserved histidine and cysteine residues (H447, H449, C455, H459, C467 and C469) in the cytoplasmic domain of JUNV GP2 as essential for the association with SSP. In a later work, the authors demonstrated that H459, C467 and C469 formed a zinc-binding structural domain that incorporated with C57

in SSP to retain SSP in GPC (Briknarova et al., 2011).

The interaction between SSP and GP2 provides a target for drug design against mammarenaviruses. Compound ST-294, showing ability to inhibit NW mammarenavirus, acts through interaction with MPER and transmembrane domain of GP2 (Bolken et al., 2006). Similarly, some mammarenavirus entry inhibitors, such as ST-193, ST-161 and 16G8, demonstrate an antiviral effect by targeting the SSP-G2 interface and perturbing the conformational rearrangement of GPC. The major mechanisms for action of these compounds seem to be stabilization of the prefusion GPC complex (Larson et al., 2008; Lee et al., 2008; York et al., 2008; Cashman et al., 2011). Ngo *et al.* (2015) reported a compound named F3406 exhibiting strong anti-LCMV activity with no cytotoxicity. The target identification revealed that residue M437 within the transmembrane domain of GP2, which has been implicated in SSP-GP2 interactions, was critical for virus susceptibility to F3406. Spence *et al.* (2014) reported a Pichindé virus GP2-derived 19-mer peptide (aa 348–366), AVP-p, with antiviral activity against OW and NW arenaviruses with a 50% inhibitory dose of 7 μ M. AVP-p was shown to induce membrane perturbation by changing the lipid organization and affecting the interaction between GPC subunits.

GLYCOSYLATION AFFECTS GPC FUNCTION AND ANTIBODY SENSITIVITY

Mammarenavirus GPC is a heavily glycosylated protein. It has been estimated that *N*-linked glycosylation accounts for 40% of the mass of GTOV GP2 (Parsy et al., 2013). *N*-Linked glycosylation plays an important role in mammarenavirus GPC expression, maturation, receptor binding, membrane fusion and immune evasion (Wright et al., 1989; Braakman and van Anken, 2000; Eschli et al., 2006; Bonhomme et al., 2011; Bonhomme et al., 2013). The GP1 of mammarenaviruses contains 4–11 predicted *N*-glycosylation sites, in which the positions of potential *N*-glycosylation sites vary considerably, as shown in Table 1. In contrast to GP1, *N*-glycosylation in GP2 is well conserved. There are four *N*-glycosylation sites that are highly conserved in GP2 of mammarenavirus, except for Lunk virus and LCMV, which lack the second *N*-glycosylation site, and Latino virus, which lacks the third. Interesting, addition of *N*-glycan in the missing glycosylation site of LCMV GP2 successfully rescued recombinant LCMV, although the virus exhibited mild viral fitness modification toward neurons (Bonhomme et al., 2013). Moreover, some NW mammarenaviruses have a unique *N*-glycosylation site located in the “top” half of the postfusion crystal structure of the GP2 trimer, while the four conserved sites are located in the “bottom” half (Igonet et al., 2011; Parsy et al., 2013), which might be the result of evolution under

Table 1. *N*-Linked glycosylation location in mammarenavirus GPC sequences. Predicted *N*-linked glycosylation was determined according to NX(S/T) motifs except for NP(S/T) using the NetNGlyc 1.0 Server (<http://www.cbs.dtu.dk/services/NetNGlyc/>)

Virus	GP1														GP1 ^a GP2				GP2 ^b Length				
Lujo	–	73	–	–	93	104	–	–	112	–	148	–	–	–	194	6	–	327	335	352	357	4	454
Dandenong	–	80	85	95	–	114	–	–	124	–	–	171	–	–	232	7	–	371	–	396	401	3	498
LCMV	–	–	85	95	–	114	–	–	124	–	–	171	–	–	232	6	–	371	–	396	401	3	498
Lunk	–	77	82	92	–	111	–	–	121	–	–	168	–	–	229	6	–	368	–	393	398	3	495
Merino walk	–	73	82	92	–	102	–	–	112	122	164	170	–	–	230	9	–	371	379	396	401	4	497
OW Ippy	66	–	78	88	97	107	–	–	117	–	159	165	197	–	228	10	–	369	377	394	399	4	495
LASSA	–	–	79	89	99	109	–	–	119	–	–	167	–	–	224	7	–	365	373	390	395	4	491
Morogoro	–	–	78	88	98	108	–	–	118	–	–	166	–	–	222	7	–	363	371	388	393	4	489
Moipeia	–	–	78	88	98	108	–	–	118	–	–	166	–	–	222	7	–	363	371	388	393	4	489
Luna	–	–	78	88	98	108	–	–	118	–	–	166	–	–	224	7	–	365	373	390	395	4	491
Mobala	–	–	78	88	98	108	–	–	118	–	–	166	–	–	224	7	–	365	373	390	395	4	491
NW Oliveros	–	75	–	90	101	112	117	–	131	–	–	180	–	234	251	9	–	389	397	414	419	4	518
C Latino	–	75	–	90	101	112	117	–	131	138	–	180	–	231	248	10	–	386	394	–	416	3	515
Junín	–	–	–	95	105	–	–	–	–	–	166	178	–	–	–	4	–	357	365	382	387	4	485
Machupo	–	–	83	95	–	–	–	–	–	137	166	178	–	–	–	5	–	368	376	393	398	4	496
Tacaribé	–	–	83	95	–	–	–	–	124	129	–	176	188	–	–	6	–	367	375	392	397	4	495
NW Cupixi	–	–	–	–	88	99	–	–	125	–	–	174	–	–	214	5	315	352	360	377	382	5	480
B Amaparí	–	–	–	–	88	99	–	–	128	–	–	174	–	–	214	5	315	352	360	377	382	5	481
Guanarito	–	–	–	–	88	–	–	–	125	–	–	174	–	202	214	5	314	351	359	376	381	5	479
Chapare	–	69	–	88	89	–	–	–	125	–	171	178	–	–	218	7	–	356	364	381	386	4	484
Sabiá	–	69	–	88	89	–	–	–	125	–	171	178	–	–	222	7	–	360	368	385	390	4	488
Pichindé	67	74	–	89	100	111	116	121	132	–	–	181	217	–	241	11	–	379	387	404	409	4	503
Pirital	–	75	–	90	101	112	117	122	133	–	–	181	218	–	243	10	–	381	389	406	411	4	510
NW A Paraná	–	74	–	89	–	111	116	119	130	–	–	170	215	219	240	10	–	378	386	403	408	4	507
Flexal	–	74	–	89	–	111	116	–	130	–	–	179	–	223	240	8	–	378	386	403	408	4	507
Allpahuayo	–	74	–	89	–	111	–	119	130	–	–	179	–	223	240	8	–	378	386	403	408	4	507
Whitewater arroyo	–	73	–	88	–	–	–	123	126	–	165	176	–	–	215	7	315	352	360	377	382	5	480
NW Skinner tank	–	73	–	88	–	–	–	–	129	–	168	178	–	–	214	6	316	353	361	378	383	5	481
D Tamiami	–	73	–	88	–	–	–	117	128	–	–	179	–	–	218	6	320	357	365	382	387	5	485
Bear canyon	–	73	–	88	–	–	–	–	130	–	–	179	–	–	216	5	–	355	363	380	385	4	483

Note: ^a The number of *N*-Linked glycosylation on GP1. ^b The number of *N*-Linked glycosylation on GP2.

selection pressure.

Deletion of the first two *N*-glycosylation sites on GP1 of LASV or LCMV results in a reduction of GPC expression, as well as the inhibition of downstream functions (Schwarz and Aebi, 2011; Bonhomme et al., 2013). The second site is well conserved across NW and OW viruses. According to the MACV GP1 conformation, the pyranose ring of the second *N*-acetylglucosamine is suggested to form a stacking interaction with F98, which is involved in

the interaction with hTfR1 (Bowden et al., 2009; Abraham et al., 2010). In LASV GP1, the second *N*-glycosylation site (N89) also contributes to building the receptor-binding cluster centered on a unique triad of histidines (Cohen-dvashi et al., 2015). Treatment of MACV GP1 with endoglycosidase F1 makes the deglycosylation GP precipitate, suggesting that *N*-glycan renders these hydrophobic molecules hydrophilic and thus solubilizes the protein (Parsy et al., 2013). It can be concluded that *N*-glycan

might solubilize the GP and facilitate the receptor recognition and binding.

Moreover, heavy glycan shielding can mask the epitope and promote nAb evasion. Antibody responses against LASV, as well as against other OW mammarenavirus, are not thought to facilitate clearance of viruses, as high viral load and high antibody titers often coexist (Johnson et al., 1987; Gunther et al., 2000; ter Meulen et al., 2000; Fisher-hoch and McCormick, 2001; Pushko et al., 2001; Meulen et al., 2004). Treatment with human convalescent plasma containing high antibodies titers shows no protective effect against LASV infection (McCormick et al., 1986). Although GP1 shedding and conformational changes were thought to be an evasion strategy (Eschli et al., 2007; Branco et al., 2010; Cohen-Dvashi et al., 2015), a recent study has shown that it was the arenavirus sugar coat that makes the nAb response inefficient (Sommerstein et al., 2015). As shown in Table 1, GP1 of NW clade B viruses have four to seven glycans. It has been reported that there was a negative correlation between nAb response and glycan density in clade B viruses (Sommerstein et al., 2015). JUNV with four glycans in GP1 generates higher nAb titers than viruses with more glycans in their GP1. Second, partially deglycosylated GP1 variants in both NW and OW viruses elicits an accelerated and more potent nAb response to the deglycosylated immunogen (Sommerstein et al., 2015). Meanwhile, antibody induced by wild-type virus neutralizes these specific glycan deletion mutations effectively (Sommerstein et al., 2015). Some key glycosylation sites, such as N99 and N119 in LASV GP1 and N373 in LASV GP2, are suggested to shield the neutralization epitopes, leading to reduction of antibody access to virus (Weber and Buchmeier, 1988; Wright et al., 1989; Bonhomme et al., 2013). It has been confirmed recently that most neutralizing human monoclonal antibodies target the linear epitope in GP1 and conformational epitopes in the GP1–GP2 interface, which are located on the glycosylated side that is not fully covered by glycosylation (Figure 3B) (Robinson et al., 2016). Vaccine development for other viruses with such a “glycan shield” provides lessons for mammarenaviruses antibody design. For example, isolation of glycan-binding broadly neutralizing antibodies from human immunodeficiency virus type 1-infected patients strongly highlights the glycan on the envelope GP as a feasible target for vaccine design (Pejchal et al., 2011; Walker et al., 2011; Mouquet et al., 2012; Kong et al., 2013; Garces et al., 2014; Garces et al., 2015).

CONCLUSION

The mammarenaviruses GPC is unique, as SSP retains and forms a heterotrimer with GP1 and GP2. The three subunits then make the (SSP/GP1/GP2)₃ homotrimer,

which is presented at the surface of the virion and is responsible for cell entry. To date, many questions based on the GPC trimer structure remain unanswered, including dissection of the interaction between SSP and GP2, and its role in virion maturation and membrane fusion, as well as elucidation of the molecular mechanism by which glycans facilitate immune evasion.

The limited treatment strategies against mammarenaviruses make the development of antiviral drugs an urgent need. The high sequence and structural homology of SSP and GP2, as well as use of the same receptor by pathogenic OW or NW viruses, provide novel targets for drug design. Investigation of the structural basis and molecular mechanisms underlying these processes will aid in the development of antiviral therapeutic strategies.

COMPLIANCE WITH ETHICS GUIDELINES

The authors declare that they have no conflict of interest. This article does not contain any studies with human or animal subjects performed by any of the authors.

REFERENCES

- Abraham J, Corbett KD, Farzan M, Choe H, Harrison SC. 2010. Structural basis for receptor recognition by New World hemorrhagic fever arenaviruses. *Nat Struct Mol Biol*, 17: 438–444.
- Abraham J, Kwong JA, Albarino CG, Lu JG, Radoshitzky SR, Salazar-Bravo J, Farzan M, Spiropoulou CF, Choe H. 2009. Host-species transferrin receptor 1 orthologs are cellular receptors for nonpathogenic new world clade B arenaviruses. *PLoS Pathog*, 5: e1000358.
- Agnihothram SS, York J, Nunberg JH. 2006. Role of the stable signal peptide and cytoplasmic domain of G2 in regulating intracellular transport of the Junin virus envelope glycoprotein complex. *J Virol*, 80: 5189–5198.
- Agnihothram SS, York J, Trahey M, Nunberg JH. 2007. Bitopic membrane topology of the stable signal peptide in the tripartite Junin virus GP-C envelope glycoprotein complex. *J Virol*, 81: 4331–4337.
- Albarino CG, Bird BH, Chakrabarti AK, Dodd KA, Flint M, Bergeron E, White DM, Nichol ST. 2011a. The major determinant of attenuation in mice of the Candid1 vaccine for Argentine hemorrhagic fever is located in the G2 glycoprotein transmembrane domain. *J Virol*, 85: 10404–10408.
- Albarino CG, Bird BH, Chakrabarti AK, Dodd KA, White DM, Bergeron E, Shrivastava-Ranjan P, Nichol ST. 2011b. Reverse genetics generation of chimeric infectious Junin/Lassa virus is dependent on interaction of homologous glycoprotein stable signal peptide and G2 cytoplasmic domains. *J Virol*, 85: 112–122.
- Armstrong C, Lillie RD. 1934. Experimental lymphocytic choriomeningitis of monkeys and mice produced by a virus encountered in studies of the 1933 St Louis encephalitis epidemic. *Public Health Rep.*, 49: 1019–1027.
- Bederka LH, Bonhomme CJ, Ling EL, Buchmeier MJ. 2014. Arenavirus stable signal peptide is the keystone subunit for glycoprotein complex organization. *MBio*, 5: e02063.
- Bedossa P, Ferlicot S, Paradis V, Dargere D, Bonvoust F, Vidaud M. 2002. Dystroglycan expression in hepatic stellate cells: role in liver fibrosis. *Lab Invest*, 82: 1053–1061.

- Beyer WR, Popplau D, Garten W, von Laer D, Lenz O. 2003. Endoproteolytic processing of the lymphocytic choriomeningitis virus glycoprotein by the subtilase SKI-1/S1P. *J Virol*, 77: 2866–2872.
- Bolken TC, Laquerre S, Zhang Y, Bailey TR, Pevear DC, Kickner SS, Sperzel LE, Jones KF, Warren TK, Amanda Lund S, Kirkwood-Watts DL, King DS, Shurtleff AC, Guttieri MC, Deng Y, Bleam M, Hruba DE. 2006. Identification and characterization of potent small molecule inhibitor of hemorrhagic fever New World arenaviruses. *Antiviral Res*, 69: 86–97.
- Bonhomme CJ, Capul AA, Lauron EJ, Bederka LH, Knopp KA, Buchmeier MJ. 2011. Glycosylation modulates arenavirus glycoprotein expression and function. *Virology*, 409: 223–233.
- Bonhomme CJ, Knopp KA, Bederka LH, Angelini MM, Buchmeier MJ. 2013. LCMV glycosylation modulates viral fitness and cell tropism. *PLoS One*, 8: e53273.
- Borrow P, Oldstone MB. 1994. Mechanism of lymphocytic choriomeningitis virus entry into cells. *Virology*, 198: 1–9.
- Bowden TA, Crispin M, Graham SC, Harvey DJ, Grimes JM, Jones EY, Stuart DI. 2009. Unusual molecular architecture of the machupo virus attachment glycoprotein. *J Virol*, 83: 8259–8265.
- Bowen MD, Peters CJ, Nichol ST. 1996. The phylogeny of New World (Tacaribe complex) arenaviruses. *Virology*, 219: 285–290.
- Bowen MD, Peters CJ, Nichol ST. 1997. Phylogenetic analysis of the Arenaviridae: patterns of virus evolution and evidence for co-speciation between arenaviruses and their rodent hosts. *Mol Phylogenet Evol*, 8: 301–316.
- Braakman I, van Anken E. 2000. Folding of viral envelope glycoproteins in the endoplasmic reticulum. *Traffic*, 1: 533–539.
- Branco LM, Grove JN, Moses LM, Goba A, Fullah M, Momoh M, Schoepp RJ, Bausch DG, Garry RF. 2010. Shedding of soluble glycoprotein 1 detected during acute Lassa virus infection in human subjects. *Virol J*, 7: 306.
- Briknarova K, Thomas CJ, York J, Nunberg JH. 2011. Structure of a zinc-binding domain in the Junin virus envelope glycoprotein. *J Biol Chem*, 286: 1528–1536.
- Buchmeier MJ, de la Torre JC, Peters CJ. 2007. *Fields Virology*, 4th edn. Philadelphia: Lippincott-Raven, pp. 1791–1828.
- Burns JW, Buchmeier MJ. 1991. Protein-protein interactions in lymphocytic choriomeningitis virus. *Virology*, 183: 620–629.
- Burri DJ, da Palma JR, Seidah NG, Zanotti G, Cendron L, Pasquato A, Kunz S. 2013. Differential recognition of Old World and New World arenavirus envelope glycoproteins by subtilisin kexin isozyme 1 (SKI-1)/site 1 protease (S1P). *J Virol*, 87: 6406–6414.
- Burri DJ, Pasqual G, Rochat C, Seidah NG, Pasquato A, Kunz S. 2012. Molecular characterization of the processing of arenavirus envelope glycoprotein precursors by subtilisin kexin isozyme-1/site-1 protease. *J Virol*, 86: 4935–4946.
- Cajimat MN, Milazzo ML, Rollin PE, Nichol ST, Bowen MD, Ksiazek TG, Fulhorst CF. 2009. Genetic diversity among Bolivian arenaviruses. *Virus Res*, 140: 24–31.
- Cao W, Henry MD, Borrow P, Yamada H, Elder JH, Ravkov EV, Nichol ST, Compans RW, Campbell KP, Oldstone MB. 1998. Identification of alpha-dystroglycan as a receptor for lymphocytic choriomeningitis virus and Lassa fever virus. *Science*, 282: 2079–2081.
- Cashman KA, Smith MA, Twenhafel NA, Larson RA, Jones KF, Allen RD, 3rd, Dai D, Chinsangaram J, Bolken TC, Hruba DE, Amberg SM, Hensley LE, Guttieri MC. 2011. Evaluation of Lassa antiviral compound ST-193 in a guinea pig model. *Antiviral Res*, 90: 70–79.
- Charrel RN, Feldmann H, Fulhorst CF, Khelifa R, de Chesse R, de Lamballerie X. 2002. Phylogeny of New World arenaviruses based on the complete coding sequences of the small genomic segment identified an evolutionary lineage produced by intrasegmental recombination. *Biochem Biophys Res Commun*, 296: 1118–1124.
- Clegg JC. 2002. Molecular phylogeny of the arenaviruses. *Curr Top Microbiol Immunol*, 262: 1–24.
- Cohen-Dvashi H, Cohen N, Israeli H, Diskin R. 2015. Molecular Mechanism for LAMP1 Recognition by Lassa Virus. *J Virol*, 89: 7584–7592.
- Cosset FL, Marianneau P, Verney G, Gallais F, Tordo N, Pecheur EI, ter Meulen J, Deubel V, Bartosch B. 2009. Characterization of Lassa virus cell entry and neutralization with Lassa virus pseudoparticles. *J Virol*, 83: 3228–3237.
- da Palma JR, Burri DJ, Oppliger J, Salamina M, Cendron L, de Laureto PP, Seidah NG, Kunz S, Pasquato A. 2014. Zymogen activation and subcellular activity of subtilisin kexin isozyme 1/site 1 protease. *J Biol Chem*, 289: 35743–35756.
- Di Simone C, Zandonatti MA, Buchmeier MJ. 1994. Acidic pH triggers LCMV membrane fusion activity and conformational change in the glycoprotein spike. *Virology*, 198: 455–465.
- Droniou-Bonzom ME, Reigner T, Oldenburg JE, Cox AU, Exline CM, Rathbun JY, Cannon PM. 2011. Substitutions in the glycoprotein (GP) of the Candid#1 vaccine strain of Junin virus increase dependence on human transferrin receptor 1 for entry and destabilize the metastable conformation of GP. *J Virol*, 85: 13457–13462.
- Eichler R, Lenz O, Garten W, Strecker T. 2006. The role of single N-glycans in proteolytic processing and cell surface transport of the Lassa virus glycoprotein GP-C. *Virol J*, 3: 41.
- Eichler R, Lenz O, Strecker T, Eickmann M, Klenk HD, Garten W. 2003a. Identification of Lassa virus glycoprotein signal peptide as a trans-acting maturation factor. *EMBO Rep*, 4: 1084–1088.
- Eichler R, Lenz O, Strecker T, Eickmann M, Klenk HD, Garten W. 2004a. Lassa virus glycoprotein signal peptide displays a novel topology with an extended endoplasmic reticulum luminal region. *J Biol Chem*, 279: 12293–12299.
- Eichler R, Lenz O, Strecker T, Garten W. 2003b. Signal peptide of Lassa virus glycoprotein GP-C exhibits an unusual length. *FEBS Lett*, 538: 203–206.
- Eichler R, Strecker T, Kolesnikova L, ter Meulen J, Weissenhorn W, Becker S, Klenk HD, Garten W, Lenz O. 2004b. Characterization of the Lassa virus matrix protein Z: electron microscopic study of virus-like particles and interaction with the nucleoprotein (NP). *Virus Res*, 100: 249–255.
- Emonet S, Lemasson JJ, Gonzalez JP, de Lamballerie X, Charrel RN. 2006. Phylogeny and evolution of old world arenaviruses. *Virology*, 350: 251–257.
- Enria DA, Barrera Oro JG. 2002. Junin virus vaccines. *Curr Top Microbiol Immunol*, 263: 239–261.
- Eschli B, Quirin K, Wepf A, Weber J, Zinkernagel R, Hengartner H. 2006. Identification of an N-terminal trimeric coiled-coil core within arenavirus glycoprotein 2 permits assignment to class I viral fusion proteins. *J Virol*, 80: 5897–5907.
- Eschli B, Zellweger RM, Wepf A, Lang KS, Quirin K, Weber J, Zinkernagel RM, Hengartner H. 2007. Early antibodies specific for the neutralizing epitope on the receptor binding subunit of the lymphocytic choriomeningitis virus glycoprotein fail to neutralize the virus. *J Virol*, 81: 11650–11657.
- Fisher-Hoch SP, McCormick JB. 2001. Towards a human Lassa fever vaccine. *Rev Med Virol*, 11: 331–341.
- Flanagan ML, Oldenburg J, Reigner T, Holt N, Hamilton GA, Martin VK, Cannon PM. 2008. New world clade B arenaviruses can use transferrin receptor 1 (TfR1)-dependent and -independent entry pathways, and glycoproteins from human pathogenic strains are associated with the use of TfR1. *J Virol*, 82: 938–948.
- Froeschke M, Basler M, Groettrup M, Dobberstein B. 2003. Long-

- lived signal peptide of lymphocytic choriomeningitis virus glycoprotein pGP-C. *J Biol Chem*, 278: 41914–41920.
- Fuller-Pace FV, Southern PJ. 1989. Detection of virus-specific RNA-dependent RNA polymerase activity in extracts from cells infected with lymphocytic choriomeningitis virus: in vitro synthesis of full-length viral RNA species. *J Virol*, 63: 1938–1944.
- Garces F, Lee JH, de Val N, Torrents de la Pena A, Kong L, Puchades C, Hua Y, Stanfield RL, Burton DR, Moore JP, Sanders RW, Ward AB, Wilson IA. 2015. Affinity Maturation of a Potent Family of HIV Antibodies Is Primarily Focused on Accommodating or Avoiding Glycans. *Immunity*, 43: 1053–1063.
- Garces F, Sok D, Kong L, McBride R, Kim HJ, Saye-Francisco KF, Julien JP, Hua Y, Cupo A, Moore JP, Paulson JC, Ward AB, Burton DR, Wilson IA. 2014. Structural evolution of glycan recognition by a family of potent HIV antibodies. *Cell*, 159: 69–79.
- Garcin D, Kolakofsky D. 1990. A novel mechanism for the initiation of Tacaribe arenavirus genome replication. *J Virol*, 64: 6196–6203.
- Goncalves AR, Moraz ML, Pasquato A, Helenius A, Lozach PY, Kunz S. 2013. Role of DC-SIGN in Lassa virus entry into human dendritic cells. *J Virol*, 87: 11504–11515.
- Gunther S, Emmerich P, Laue T, Kuhle O, Asper M, Jung A, Grewing T, ter Meulen J, Schmitz H. 2000. Imported lassa fever in Germany: molecular characterization of a new lassa virus strain. *Emerg Infect Dis*, 6: 466–476.
- Hara Y, Kanagawa M, Kunz S, Yoshida-Moriguchi T, Satz JS, Kobayashi YM, Zhu Z, Burden SJ, Oldstone MB, Campbell KP. 2011. Like-acetylglycosaminyltransferase (LARGE)-dependent modification of dystroglycan at Thr-317/319 is required for laminin binding and arenavirus infection. *Proc Natl Acad Sci U S A*, 108: 17426–17431.
- Harrison SC. 2008. Viral membrane fusion. *Nat Struct Mol Biol*, 15: 690–698.
- Hastie KM, Igonet S, Sullivan BM, Legrand P, Zandonatti MA, Robinson JE, Garry RF, Rey FA, Oldstone MB, Saphire EO. 2016. Crystal structure of the prefusion surface glycoprotein of the prototypic arenavirus LCMV. *Nat Struct Mol Biol*, 23: 513–521.
- Igonet S, Vaney MC, Vonnrhein C, Bricogne G, Stura EA, Hengartner H, Eschli B, Rey FA. 2011. X-ray structure of the arenavirus glycoprotein GP2 in its postfusion hairpin conformation. *Proc Natl Acad Sci U S A*, 108: 19967–19972.
- Imperiali M, Thoma C, Pavoni E, Brancaccio A, Callewaert N, Oxenius A. 2005. O Mannosylation of alpha-dystroglycan is essential for lymphocytic choriomeningitis virus receptor function. *J Virol*, 79: 14297–14308.
- Jae LT, Raaben M, Herbert AS, Kuehne AI, Wirchnianski AS, Soh TK, Stubbs SH, Janssen H, Damme M, Saftig P, Whelan SP, Dye JM, Brummelkamp TR. 2014. Lassa virus entry requires a trigger-induced receptor switch. *Science*, 344: 1506–1510.
- Jae LT, Raaben M, Riemersma M, van Beusekom E, Blomen VA, Velds A, Kerkhoven RM, Carette JE, Topaloglu H, Meinecke P, Wessels MW, Lefeber DJ, Whelan SP, van Bokhoven H, Brummelkamp TR. 2013. Deciphering the glycosylome of dystroglycanopathies using haploid screens for lassa virus entry. *Science*, 340: 479–483.
- Johnson KM, McCormick JB, Webb PA, Smith ES, Elliott LH, King IJ. 1987. Clinical virology of Lassa fever in hospitalized patients. *J Infect Dis*, 155: 456–464.
- Klewitz C, Klenk HD, ter Meulen J. 2007. Amino acids from both N-terminal hydrophobic regions of the Lassa virus envelope glycoprotein GP-2 are critical for pH-dependent membrane fusion and infectivity. *J Gen Virol*, 88: 2320–2328.
- Kong L, Lee JH, Doores KJ, Murin CD, Julien JP, McBride R, Liu Y, Marozsan A, Cupo A, Klasse PJ, Hoffenberg S, Caulfield M, King CR, Hua Y, Le KM, Khayat R, Deller MC, Clayton T, Tien H, Feizi T, Sanders RW, Paulson JC, Moore JP, Stanfield RL, Burton DR, Ward AB, Wilson IA. 2013. Supersite of immune vulnerability on the glycosylated face of HIV-1 envelope glycoprotein gp120. *Nat Struct Mol Biol*, 20: 796–803.
- Kranzusch PJ, Whelan SP. 2012. Architecture and regulation of negative-strand viral enzymatic machinery. *RNA Biol*, 9: 941–948.
- Kunz S, Edelman KH, de la Torre JC, Gorney R, Oldstone MB. 2003. Mechanisms for lymphocytic choriomeningitis virus glycoprotein cleavage, transport, and incorporation into virions. *Virology*, 314: 168–178.
- Kunz S, Rojek JM, Kanagawa M, Spiropoulou CF, Barresi R, Campbell KP, Oldstone MB. 2005. Posttranslational modification of alpha-dystroglycan, the cellular receptor for arenaviruses, by the glycosyltransferase LARGE is critical for virus binding. *J Virol*, 79: 14282–14296.
- Landau M, Mayrose I, Rosenberg Y, Glaser F, Martz E, Pupko T, Ben-Tal N. 2005. ConSurf 2005: the projection of evolutionary conservation scores of residues on protein structures. *Nucleic Acids Res*, 33: W299–W302.
- Larson RA, Dai D, Hosack VT, Tan Y, Bolken TC, Hruby DE, Amberg SM. 2008. Identification of a broad-spectrum arenavirus entry inhibitor. *J Virol*, 82: 10768–10775.
- Lee AM, Rojek JM, Spiropoulou CF, Gundersen AT, Jin W, Shaginian A, York J, Nunberg JH, Boger DL, Oldstone MB, Kunz S. 2008. Unique small molecule entry inhibitors of hemorrhagic fever arenaviruses. *J Biol Chem*, 283: 18734–18742.
- Lenz O, ter Meulen J, Klenk HD, Seidah NG, Garten W. 2001. The Lassa virus glycoprotein precursor GP-C is proteolytically processed by subtilase SKI-1/S1P. *Proc Natl Acad Sci U S A*, 98: 12701–12705.
- Leung WC, Leung MF, Rawls WE. 1979. Distinctive RNA transcriptase, polyadenylic acid polymerase, and polyuridylic acid polymerase activities associated with Pichinde virus. *J Virol*, 30: 98–107.
- Li S, Sun Z, Pryce R, Parsy ML, Fehling SK, Schlie K, Siebert CA, Garten W, Bowden TA, Strecker T, Huiskenon JT. 2016. Acidic pH-Induced Conformations and LAMP1 Binding of the Lassa Virus Glycoprotein Spike. *PLoS Pathog*, 12: e1005418.
- Maiztegui JI, McKee KT, Jr., Barrera Oro JG, Harrison LH, Gibbs PH, Feuillade MR, Enria DA, Briggiler AM, Levis SC, Ambrosio AM, Halsey NA, Peters CJ. 1998. Protective efficacy of a live attenuated vaccine against Argentine hemorrhagic fever. *AHF Study Group. J Infect Dis*, 177: 277–283.
- Martinez MG, Bialecki MA, Belouard S, Cordo SM, Candurra NA, Whittaker GR. 2013. Utilization of human DC-SIGN and L-SIGN for entry and infection of host cells by the New World arenavirus, Junin virus. *Biochem Biophys Res Commun*, 441: 612–617.
- Martinez MG, Cordo SM, Candurra NA. 2007. Characterization of Junin arenavirus cell entry. *J Gen Virol*, 88: 1776–1784.
- McCormick JB, King IJ, Webb PA, Scribner CL, Craven RB, Johnson KM, Elliott LH, Belmont-Williams R. 1986. Lassa fever. Effective therapy with ribavirin. *N Engl J Med*, 314: 20–26.
- Messina EL, York J, Nunberg JH. 2012. Dissection of the role of the stable signal peptide of the arenavirus envelope glycoprotein in membrane fusion. *J Virol*, 86: 6138–6145.
- Meulen J, Badusche M, Satoguina J, Strecker T, Lenz O, Loeliger C, Sakho M, Koulemou K, Koivogui L, Hoerauf A. 2004. Old and New World arenaviruses share a highly conserved epitope in the fusion domain of the glycoprotein 2, which is recognized by Lassa virus-specific human CD4+ T-cell clones. *Virology*, 321: 134–143.
- Meyer BJ, de la Torre JC, Southern PJ. 2002. Arenaviruses: genomic

- RNAs, transcription, and replication. *Curr Top Microbiol Immunol*, 262: 139–157.
- Mouquet H, Scharf L, Euler Z, Liu Y, Eden C, Scheid JF, Halper-Stromberg A, Gnanapragasam PN, Spencer DI, Seaman MS, Schuitemaker H, Feizi T, Nussenzweig MC, Bjorkman PJ. 2012. Complex-type N-glycan recognition by potent broadly neutralizing HIV antibodies. *Proc Natl Acad Sci U S A*, 109: E3268–3277.
- Ngo N, Cubitt B, Iwasaki M, de la Torre JC. 2015. Identification and Mechanism of Action of a Novel Small-Molecule Inhibitor of Arenavirus Multiplication. *J Virol*, 89: 10924–10933.
- Oldstone MB. 2007. A suspenseful game of 'hide and seek' between virus and host. *Nat Immunol*, 8: 325–327.
- Parsy ML, Harlos K, Huiskonen JT, Bowden TA. 2013. Crystal structure of Venezuelan hemorrhagic fever virus fusion glycoprotein reveals a class I postfusion architecture with extensive glycosylation. *J Virol*, 87: 13070–13075.
- Pasqual G, Rojek JM, Masin M, Chatton JY, Kunz S. 2011. Old world arenaviruses enter the host cell via the multivesicular body and depend on the endosomal sorting complex required for transport. *PLoS Pathog*, 7: e1002232.
- Pasquato A, Burri DJ, Traba EG, Hanna-El-Daher L, Seidah NG, Kunz S. 2011. Arenavirus envelope glycoproteins mimic autoprocessing sites of the cellular proprotein convertase subtilisin kexin isozyme-1/site-1 protease. *Virology*, 417: 18–26.
- Pejchal R, Doores KJ, Walker LM, Khayat R, Huang PS, Wang SK, Stanfield RL, Julien JP, Ramos A, Crispin M, Depetris R, Katpally U, Marozsan A, Cupo A, Malveste S, Liu Y, McBride R, Ito Y, Sanders RW, Ogohara C, Paulson JC, Feizi T, Scanlan CN, Wong CH, Moore JP, Olson WC, Ward AB, Poignard P, Schief WR, Burton DR, Wilson IA. 2011. A potent and broad neutralizing antibody recognizes and penetrates the HIV glycan shield. *Science*, 334: 1097–1103.
- Perez M, Craven RC, de la Torre JC. 2003. The small RING finger protein Z drives arenavirus budding: implications for antiviral strategies. *Proc Natl Acad Sci U S A*, 100: 12978–12983.
- Pinschewer DD, Perez M, de la Torre JC. 2003. Role of the virus nucleoprotein in the regulation of lymphocytic choriomeningitis virus transcription and RNA replication. *J Virol*, 77: 3882–3887.
- Pushko P, Geisbert J, Parker M, Jahrling P, Smith J. 2001. Individual and bivalent vaccines based on alphavirus replicons protect guinea pigs against infection with Lassa and Ebola viruses. *J Virol*, 75: 11677–11685.
- Quirin K, Eschli B, Scheu I, Poort L, Kartenbeck J, Helenius A. 2008. Lymphocytic choriomeningitis virus uses a novel endocytic pathway for infectious entry via late endosomes. *Virology*, 378: 21–33.
- Radoshitzky SR, Abraham J, Spiropoulou CF, Kuhn JH, Nguyen D, Li W, Nagel J, Schmidt PJ, Nunberg JH, Andrews NC, Farzan M, Choe H. 2007. Transferrin receptor 1 is a cellular receptor for New World haemorrhagic fever arenaviruses. *Nature*, 446: 92–96.
- Radoshitzky SR, Bao Y, Buchmeier MJ, Charrel RN, Clawson AN, Clegg CS, DeRisi JL, Emonet S, Gonzalez JP, Kuhn JH, Lukashevich IS, Peters CJ, Romanowski V, Salvato MS, Stenglein MD, de la Torre JC. 2015. Past, present, and future of arenavirus taxonomy. *Arch Virol*, 160: 1851–1874.
- Radoshitzky SR, Longobardi LE, Kuhn JH, Retterer C, Dong L, Clester JC, Kota K, Carra J, Bavari S. 2011. Machupo virus glycoprotein determinants for human transferrin receptor 1 binding and cell entry. *PLoS One*, 6: e21398.
- Robert X, Gouet P. 2014. Deciphering key features in protein structures with the new ENDscript server. *Nucleic Acids Res*, 42: W320–W324.
- Robinson JE, Hastie KM, Cross RW, Yenni RE, Elliott DH, Rouelle JA, Kannadka CB, Smira AA, Garry CE, Bradley BT, Yu H, Shaffer JG, Boisen ML, Hartnett JN, Zandonatti MA, Rowland MM, Heinrich ML, Martinez-Sobrido L, Cheng B, de la Torre JC, Andersen KG, Goba A, Momoh M, Fullah M, Gbakie M, Kanneh L, Koroma VJ, Fonnier R, Jalloh SC, Kargbo B, Vandi MA, Gbetuwa M, Ikponmwoosa O, Asogun DA, Okokhere PO, Follarin OA, Schieffelin JS, Pitts KR, Geisbert JB, Kulakowski PC, Wilson RB, Happi CT, Sabeti PC, Gevao SM, Khan SH, Grant DS, Geisbert TW, Saphire EO, Branco LM, Garry RF. 2016. Most neutralizing human monoclonal antibodies target novel epitopes requiring both Lassa virus glycoprotein subunits. *Nat Commun*, 7: 11544.
- Rojek JM, Campbell KP, Oldstone MB, Kunz S. 2007a. Old World arenavirus infection interferes with the expression of functional alpha-dystroglycan in the host cell. *Mol Biol Cell*, 18: 4493–4507.
- Rojek JM, Kunz S. 2008. Cell entry by human pathogenic arenaviruses. *Cell Microbiol*, 10: 828–835.
- Rojek JM, Lee AM, Nguyen N, Spiropoulou CF, Kunz S. 2008a. Site 1 protease is required for proteolytic processing of the glycoproteins of the South American hemorrhagic fever viruses Junin, Machupo, and Guanarito. *J Virol*, 82: 6045–6051.
- Rojek JM, Sanchez AB, Nguyen NT, de la Torre JC, Kunz S. 2008b. Different mechanisms of cell entry by human-pathogenic Old World and New World arenaviruses. *J Virol*, 82: 7677–7687.
- Rojek JM, Spiropoulou CF, Campbell KP, Kunz S. 2007b. Old World and clade C New World arenaviruses mimic the molecular mechanism of receptor recognition used by alpha-dystroglycan's host-derived ligands. *J Virol*, 81: 5685–5695.
- Salvato MS, Clegg JCS, Buchmeier MJ, Charrel RN, Gonzalez JP, Lukashevich IS, Peters CJ, Rico-Hesse R, Romanowski V. 2005. Family Arenaviridae. In: *Virus Taxonomy, Eighth report of the International Committee on Taxonomy of Viruses*. Van Regenmortel MHV, Fauquet CM, Mayo MA, Maniloff J, Desselberger U, Ball LA (Eds.). Cambridge: Academic Press.
- Salvato MS, Schweighofer KJ, Burns J, Shimomaye EM. 1992. Biochemical and immunological evidence that the 11 kDa zinc-binding protein of lymphocytic choriomeningitis virus is a structural component of the virus. *Virus Res*, 22: 185–198.
- Salvato MS, Shimomaye EM. 1989. The completed sequence of lymphocytic choriomeningitis virus reveals a unique RNA structure and a gene for a zinc finger protein. *Virology*, 173: 1–10.
- Saunders AA, Ting JP, Meisner J, Neuman BW, Perez M, de la Torre JC, Buchmeier MJ. 2007. Mapping the landscape of the lymphocytic choriomeningitis virus stable signal peptide reveals novel functional domains. *J Virol*, 81: 5649–5657.
- Schmitt AP, Leser GP, Morita E, Sundquist WI, Lamb RA. 2005. Evidence for a new viral late-domain core sequence, FPIV, necessary for budding of a paramyxovirus. *J Virol*, 79: 2988–2997.
- Schrepf S, Froeschke M, Giroglou T, von Laer D, Dobberstein B. 2007. Signal peptide requirements for lymphocytic choriomeningitis virus glycoprotein C maturation and virus infectivity. *J Virol*, 81: 12515–12524.
- Schwarz F, Aebi M. 2011. Mechanisms and principles of N-linked protein glycosylation. *Curr Opin Struct Biol*, 21: 576–582.
- Seregin AV, Yun NE, Miller M, Aronson J, Smith JK, Walker AG, Smith JN, Huang C, Manning JT, de la Torre JC, Paessler S. 2015. The glycoprotein precursor gene of Junin virus determines the virulence of the Romero strain and the attenuation of the Candid #1 strain in a representative animal model of Argentine hemorrhagic fever. *J Virol*, 89: 5949–5956.
- Shimajima M, Kawaoka Y. 2012. Cell Surface Molecules Involved in Infection Mediated by Lymphocytic Choriomeningitis Virus Glycoprotein. *J Vet Med Sci*, 74: 1363–1366.
- Shimajima M, Stroher U, Ebihara H, Feldmann H, Kawaoka Y.

2012. Identification of cell surface molecules involved in dystroglycan-independent Lassa virus cell entry. *J Virol*, 86: 2067–2078.
- Sommerstein R, Flatz L, Remy MM, Malinge P, Magistrelli G, Fischer N, Sahin M, Bergthaler A, Igonet S, Ter Meulen J, Rigo D, Meda P, Rabah N, Coutard B, Bowden TA, Lambert PH, Siegrist CA, Pinschewer DD. 2015. Arenavirus Glycan Shield Promotes Neutralizing Antibody Evasion and Protracted Infection. *PLoS Pathog*, 11: e1005276.
- Spence JS, Melnik LI, Badani H, Wimley WC, Garry RF. 2014. Inhibition of arenavirus infection by a glycoprotein-derived peptide with a novel mechanism. *J Virol*, 88: 8556–8564.
- Strecker T, Eichler R, Meulen J, Weissenhorn W, Dieter Klenk H, Garten W, Lenz O. 2003. Lassa virus Z protein is a matrix protein and sufficient for the release of virus-like particles [corrected]. *J Virol*, 77: 10700–10705.
- Tani H, Iha K, Shimojima M, Fukushi S, Taniguchi S, Yoshikawa T, Kawaoka Y, Nakasone N, Ninomiya H, Saijo M, Morikawa S. 2014. Analysis of Lujo virus cell entry using pseudotype vesicular stomatitis virus. *J Virol*, 88: 7317–7330.
- ter Meulen J, Badusche M, Kuhnt K, Doetze A, Satoguina J, Marti T, Loeliger C, Koulemou K, Koivogui L, Schmitz H, Fleischer B, Hoerauf A. 2000. Characterization of human CD4(+) T-cell clones recognizing conserved and variable epitopes of the Lassa virus nucleoprotein. *J Virol*, 74: 2186–2192.
- Vela EM, Zhang L, Colpitts TM, Davey RA, Aronson JF. 2007. Arenavirus entry occurs through a cholesterol-dependent, non-caveolar, clathrin-mediated endocytic mechanism. *Virology*, 369: 1–11.
- Walker LM, Huber M, Doores KJ, Falkowska E, Pejchal R, Julien JP, Wang SK, Ramos A, Chan-Hui PY, Moyle M, Mitcham JL, Hammond PW, Olsen OA, Phung P, Fling S, Wong CH, Phogat S, Wrinn T, Simek MD, Protocol GPI, Koff WC, Wilson IA, Burton DR, Pognard P. 2011. Broad neutralization coverage of HIV by multiple highly potent antibodies. *Nature*, 477: 466–470.
- Weber EL, Buchmeier MJ. 1988. Fine mapping of a peptide sequence containing an antigenic site conserved among arenaviruses. *Virology*, 164: 30–38.
- Wright KE, Salvato MS, Buchmeier MJ. 1989. Neutralizing epitopes of lymphocytic choriomeningitis virus are conformational and require both glycosylation and disulfide bonds for expression. *Virology*, 171: 417–426.
- Yamamoto T, Kato Y, Karita M, Kawaguchi M, Shibata N, Kobayashi M. 2004. Expression of genes related to muscular dystrophy with lissencephaly. *Pediatr Neurol*, 31: 183–190.
- York J, Agnihotram SS, Romanowski V, Nunberg JH. 2005. Genetic analysis of heptad-repeat regions in the G2 fusion subunit of the Junin arenavirus envelope glycoprotein. *Virology*, 343: 267–274.
- York J, Dai D, Amberg SM, Nunberg JH. 2008. pH-induced activation of arenavirus membrane fusion is antagonized by small-molecule inhibitors. *J Virol*, 82: 10932–10939.
- York J, Nunberg JH. 2006. Role of the stable signal peptide of Junin arenavirus envelope glycoprotein in pH-dependent membrane fusion. *J Virol*, 80: 7775–7780.
- York J, Nunberg JH. 2007a. Distinct requirements for signal peptidase processing and function in the stable signal peptide subunit of the Junin virus envelope glycoprotein. *Virology*, 359: 72–81.
- York J, Nunberg JH. 2007b. A novel zinc-binding domain is essential for formation of the functional Junin virus envelope glycoprotein complex. *J Virol*, 81: 13385–13391.
- York J, Nunberg JH. 2009. Intersubunit interactions modulate pH-induced activation of membrane fusion by the Junin virus envelope glycoprotein GPC. *J Virol*, 83: 4121–4126.
- York J, Romanowski V, Lu M, Nunberg JH. 2004. The signal peptide of the Junin arenavirus envelope glycoprotein is myristoylated and forms an essential subunit of the mature G1-G2 complex. *J Virol*, 78: 10783–10792.
- Young PR, Howard CR. 1983. Fine structure analysis of Pichinde virus nucleocapsids. *J Gen Virol*, 64: 833–842.