



REVIEW

Manipulation of the host cell membrane by human γ -herpesviruses EBV and KSHV for pathogenesis

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The cell membrane regulates many physiological processes including cellular communication, homing and metabolism. It is therefore not surprising that the composition of the host cell membrane is manipulated by intracellular pathogens. Among these, the human oncogenic herpesviruses Epstein–Barr virus (EBV) and Kaposi’s sarcoma-associated herpesvirus (KSHV) exploit the host cell membrane to avoid immune surveillance and promote viral replication. Accumulating evidence has shown that both EBV and KSHV directly encode several similar membrane-associated proteins, including receptors and receptor-specific ligands (cytokines and chemokines), to increase virus fitness in spite of host antiviral immune responses. These proteins are expressed individually at different phases of the EBV/KSHV life cycle and employ various mechanisms to manipulate the host cell membrane. In recent decades, much effort has been made to address how these membrane-based signals contribute to viral tumorigenesis. In this review, we summarize and highlight the recent understanding of how EBV and KSHV similarly manipulate host cell membrane signals, particularly how remodeling of the cell membrane allows EBV and KSHV to avoid host antiviral immune responses and favors their latent and lytic infection.

KEYWORDS cell membrane remodeling; Epstein-Barr virus (EBV); Kaposi’s sarcoma-associated herpesvirus (KSHV)

INTRODUCTION

Mammalian cells are encased by a selectively permeable membrane, which is composed of lipids and proteins and separates the cellular cytoplasm from its surroundings (Goldberg and Riordan, 1986). The cell membrane proteins usually contain carbohydrate residues directed towards the exterior and are important for the interaction of cells with each other and with external proteins (Goldberg and Riordan, 1986). To maintain proper

homeostasis, the protein and lipid components of the cell membrane are finely controlled in a dynamic and responsive state. It has been widely demonstrated that the external (plasma) membrane is often damaged in a wide range of human diseases, which compromises the integrity of the cells (Goldberg and Riordan, 1986). Due to the host cell membrane plays a crucial role in defense, particularly serving as an initial barrier for pathogen infection, it has been a target frequently hijacked by pathogens to invade host cells (Chakraborty et al., 2012). In virus-associated diseases, it is also very common for changes in the composition and function of proteins within the host cell membrane to occur during viral pathogenesis (Chakraborty et al., 2012).

Epstein-Barr virus (EBV or HHV-4) and Kaposi’s sarcoma-associated herpesvirus (KSHV or HHV-8) are members of the human γ -herpesvirus subfamily and are

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lymphotropic viruses in natural or experimental settings (Epstein et al., 1964; Chang et al., 1994). Like other herpesviruses, they are capable of establishing latent infection in host cells and reactivating for lytic replication under certain conditions. EBV has been linked with many diseases in humans, including infectious mononucleosis (IM), Burkitt's lymphoma (BL), nasopharyngeal carcinoma (NPC), Hodgkin's disease (HD) and T-cell lymphoma; while KSHV has been associated with Kaposi's sarcoma (KS), primary effusion lymphoma (PEL) and multicentric Castleman's disease (MCD) (Cesarman et al., 1995; Mesri et al., 2010). It has been documented that EBV can infect B cells, epithelial cells and some T cells (Spear and Longnecker, 2003; Calderwood et al., 2007). In contrast, it seems that KSHV has a broader range of cell tropism and is able to infect B cells, endothelial cells, epithelial cells and cells of monocyte or macrophage lineage *in vitro* (Veettil et al., 2014). Although the cell tropism and pathogenesis of these two oncogenic viruses differ, increasing evidence has shown that they exploit similar strategies to invade host cells during their latent and lytic life cycles, particularly in induction of host cell proliferation, transformation and tumorigenesis, and escape from host antiviral responses (Riethmüller et al., 2006).

Given the important roles of the host cell membrane in cellular metabolism, homing, communication and especially immune surveillance, it is not surprising that both EBV and KSHV have evolved specialized proteins that manipulate the composition and function of host cell membrane proteins to avoid host antiviral immune responses and establish persistent infection (Chazal and Gerlier, 2003; Heaton and Randall, 2011; Mazzon and Mercer, 2014). Deciphering how EBV and KSHV hijack the host cell membrane will help to provide new insights into understanding the mechanisms of cell transformation and oncogenesis induced by other tumor-inducing viruses. Indeed, open reading frame (ORF) analysis from viral genome sequences has revealed that both EBV and KSHV encode several important membrane-associated proteins, which include regulators of cell transformation and signaling, modulators or mimics of cellular receptors, and viral cytokines and chemokines, to directly or

indirectly take over host cell membrane signaling (Figure 1). In this review, we summarize the functional similarities of the viral proteins encoded by EBV or KSHV for manipulating the composition of host cell membranes and signaling for viral pathogenesis, and highlight the recent evidence for how EBV and KSHV similarly or differently utilize these membrane-associated proteins to facilitate their latent and lytic infection.

UNIQUE VIRAL MEMBRANE PROTEINS FOR CELL TRANSFORMATION AND SIGNALING

Increasing evidence indicates that the herpesviruses, including EBV and KSHV, usually encode several proteins for cell transformation and signaling to protect and preserve the virus-infected cells. Among these proteins are unique viral membrane proteins that have no amino acid sequence similarity to cellular proteins. For instance, Latent membrane protein 1 (LMP1) and K1 are viral oncoproteins encoded by EBV and KSHV, respectively. Although these two genes have highly divergent sequences, the function of the gene products seems to be conserved.

Extensive studies have revealed that the LMP1 protein contains six transmembrane-spanning domains and a 200 amino acid cytoplasmic domain at the carboxyl terminus (Wang et al., 1985; Wang et al., 1988). The carboxyl terminus of LMP1 can be divided into two functional motif regions—CTAR1 and CTAR2. LMP1 is expressed in latently infected B cells and can be upregulated during the lytic replication cycle in epithelial and B cells. It has been shown that LMP1 can transform fibroblasts *in vitro* and mimics the B-cell activation antigen CD40. Similar to CD40 and other receptors, CTAR1 and CTAR2 in the cytoplasmic domain of LMP1 are associated with several essential proteins including the tumor-necrosis factor (TNF) receptor-associated molecules TRAF and TRADD, for activation of the nuclear factor (NF)- κ B pathway and for EBV-induced immortalization of B lymphocytes (Izumi et al., 1997). However, distinct from CD40, the transduction of LMP1 signals is not due to extracellular ligands or cross-linking but is caused by

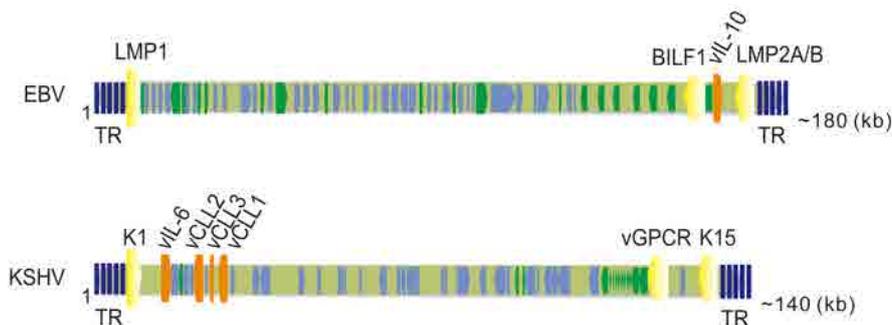


Figure 1. Genomic localization of the host cell membrane proteins, cytokines and chemokines encoded by EBV and KSHV. Host cell membrane proteins are shown in yellow; cytokines or chemokines are indicated in orange. Latency-associated genes are in green; lytic genes are in light blue. TR, terminal repeats.

multimerization of LMP1 through its transmembrane domains, which mimics ligand-induced CD40 receptor aggregation (Figure 2A). Moreover, this multimerization leads to constitutive activation of NF- κ B, cJun N-terminal kinase (JNK) and phosphoinositide 3-kinase (PI3K) activity, as well as induction of expression of many downstream genes including *Bcl2* for cell survival (Hatzivassiliou et al., 1998). Hence, LMP1 mainly functions as a B-lymphocyte CD40 receptor to contribute to EBV-infected cell growth, survival and transformation. Interestingly, recent studies in animal models have shown that EBV with LMP1 deficiency can still establish long-term viral latency *in vivo*, but is unable to induce lymphomas (Gujer et al., 2015; Ma et al., 2015), and LMP1 likely supports the survival of EBV-infected cells in a competitive environment that favors differentiation into memory B cells rather than plasma cells (Thorley-Lawson et al., 2013). Importantly, LMP1 is also able to promote cell migration and globally regulate chromatin and expression of genes including tumor suppressor DOK1 (Siouda et al., 2014), pro-inflammatory cytokines (Xiao et al., 2014), and small non-coding RNA (Motsch et al., 2007; Amort et al., 2015). The fact that LMP1 is present in exosomes from EBV-infected B cells and NPC cells indicates that LMP1 not only plays a role in immune modulation but also in cell communication and tumor microenvironment homeostasis (Meckes et al., 2010).

Similar to the LMP1 of EBV, KSHV encodes an ORF called K1 (Lagunoff and Ganem, 1997; Lee et al., 1998b). In contrast to LMP1, however, K1 is a single transmembrane glycoprotein (Figure 2B) (Zong et al., 1999). Although the amino-terminal domain of K1 is highly variable, the carboxyl-terminal cytoplasmic domain is relatively well conserved. Moreover, this carboxyl terminus contains a functional immunoreceptor tyrosine-based activation motif (ITAM), which is important for cellular activation (Lee et al., 1998a; Lagunoff et al., 1999). Interestingly, similar to LMP1, K1 appears to be constitutively activated independently of the ITAM-based signal transduction of ligand-receptor interaction and is also able to transform fibroblasts *in vitro* (Lagunoff et al., 1999). In addition, it has also been shown that K1 interacts with several cellular proteins including p85, Vav, Syk and Akt kinases to activate NF- κ B and NF-AT pathways for cell growth and B-cell activation (Bowser et al., 2002; Tomlinson and Damania, 2004; Tomlinson and Damania, 2008). In contrast with LMP1, K1 has been shown to induce angiogenesis and cell invasion through promoting the secretion of vascular endothelial growth factor (VEGF) and MMP-9 (Wang et al., 2004) and to transform endothelial cells via activation of the PI3K pathway (Wang et al., 2006), as well as functioning in anti-apoptosis and endocytosis (Tomlinson and Damania,

2008; Wen and Damania, 2010). Interestingly, Damania's group showed that K1 was also highly upregulated by the master regulator of lytic replication, RTA, and in turn augmented viral lytic replication (Bowser et al., 2006; Zhang et al., 2016).

VIRAL SIGNAL MODULATORS OF THE B-CELL RECEPTOR

The activation of immune cells through a receptor at the membrane surface is important for antiviral immune responses, and is often targeted by viral infection. During EBV latent infection, LMP2A is highly expressed in B cells for impairing B-cell activation (Fruehling and Longnecker, 1997; Caldwell et al., 1998). It has been demonstrated that LMP2A has 12 transmembrane domains and short amino-terminal and carboxyl-terminal domains (Figure 2C). There are three tyrosine-based SH2 domain binding sites located in the amino-terminal cytoplasmic region, two of which form a functional ITAM (Fruehling and Longnecker, 1997), which is required for LMP2A to associate with Lyn, Syk and Csk kinases (Burkhardt et al., 1992). In contrast to LMP1, although LMP2A is dispensable for EBV-induced immortalization of B cells, it can block normal B-cell receptor (BCR) signaling in EBV-negative B cells (Miller et al., 1993). This indicates that LMP2A may play a significant role in the establishment and maintenance of EBV latency (Miller et al., 1994; Miller et al., 1995). Many regulatory functions of LMP2A on host cell proliferation, activation, migration, transformation and survival have been reported and were recently summarized by Cen and Longnecker (2015). For instance, the recruitment of Syk and Akt kinases is required for LMP2A to transform and increase migration of epithelial cells, which relies on its ITAM motif (Lu et al., 2006; Fotheringham et al., 2012; Fukuda and Kawaguchi, 2014). In LMP2A transgenic mice, LMP2A could also activate constitutive signals for B cell survival through ITAM/Syk, Ras/PI3K/Akt and mitogen-activated protein kinase (ERK/MAPK) pathways (Merchant et al., 2000; Portis and Longnecker, 2004; Fukuda and Longnecker, 2005; Anderson and Longnecker, 2008). In addition, LMP2A can block the expression of LMP1 to indirectly inhibit interleukin (IL)-6 expression through the Janus kinase/signal transducers and activators of transcription (JAK-STAT) pathway (Stewart et al., 2004), and can drive cell progression by reducing the expression level of p27 (Fish et al., 2014). Furthermore, it has been shown that LMP2A is highly expressed in the EBV latency types II and III, including EBV-immortalized B-lymphoblastoid cell lines (LCLs), Hodgkin's and non-Hodgkin's lymphoma, other lymphoproliferative malignancies, as well as NPC and gastric carcinoma (Qu et al.,

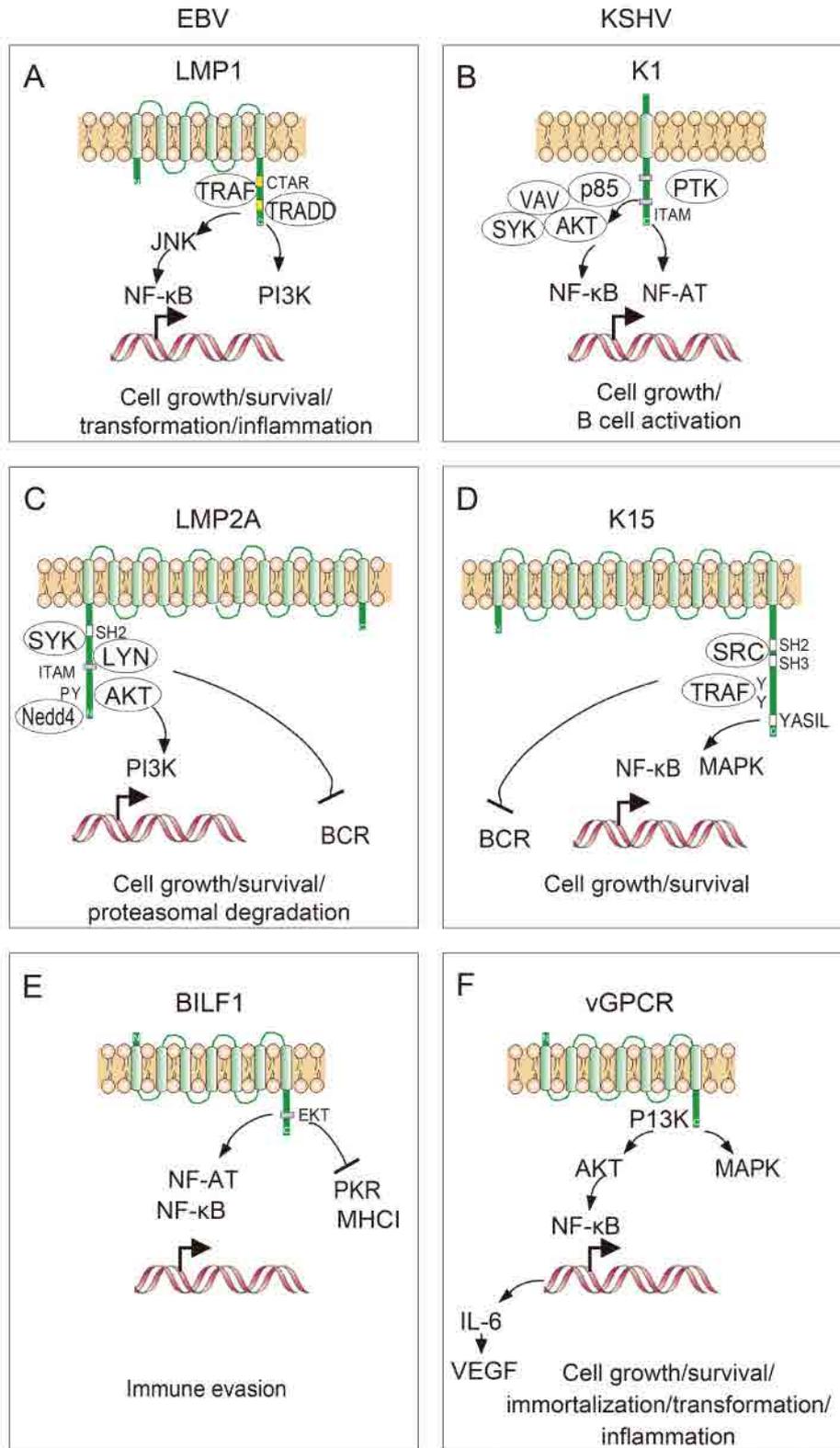


Figure 2. Structure of the host cell membrane proteins encoded by EBV and KSHV. (A–F) LMP1 and K1, LMP2A and K15, and BILF1 and vGPCR are encoded individually by EBV and KSHV, respectively. Interactions with cellular partners and activation of cellular signaling pathways are indicated.

2000; Fox et al., 2010; Han et al., 2012). These discoveries indicate that LMP2A may provide a potential therapeutic target for the treatment of EBV-associated cancers.

In the equivalent position to the EBV LMP2A, KSHV encodes a distinct ORF named K15 (Glenn et al., 1999). It has been demonstrated that K15 contains 4–12 transmembrane domains and a short amino-terminal cytoplasmic domain (Figure 2D). In contrast to LMP2A, K15 is weakly expressed in KSHV-latently infected primary effusion lymphoma (PEL) cells, while levels are significantly increased upon stimulation with chemicals such as 12-*O*-tetradecanoyl-phorbol-13-acetate (TPA). Similar to LMP2A, different variants of K15 exist; however, the signaling motifs (including SH2 and SH3 binding motifs and a YASIL sequence) located in the cytoplasmic domain of K15 are highly conserved, and the tyrosine residue within the putative SH2 binding motif is constitutively phosphorylated. Like LMP2A, K15 is capable of blocking BCR signal transduction, although it is unable to elicit cellular signal transduction upon antibody stimulation. This could explain to a certain extent why LMP2A but not K15 is highly expressed during viral latency. Interestingly, K15 was exclusively found to induce angiogenesis and invasiveness of endothelial cells (Bala et al., 2012; Gramolelli et al., 2015). In addition, it was demonstrated that several intracellular signaling pathways including MAPK and NF- κ B were activated by K15 to induce inflammation and angiogenesis (Brinkmann et al., 2003; Brinkmann et al., 2007; Wang et al., 2007; Pietrek et al., 2010).

VIRAL MIMICKERS OF THE G-PROTEIN-COUPLED RECEPTOR

In addition to encoding a viral modulator of host cell membrane signaling, EBV and KSHV also express proteins that mimic host cell membrane receptors. G-protein-coupled receptors (GPCRs) are a diverse family of membrane receptors that can be activated by a variety of ligands (Rosenbaum et al., 2009). All GPCRs contains seven transmembrane domains and couple to hetero-G-proteins in response to receptor activation (Rosenbaum et al., 2009). EBV and KSHV, as oncogenic members of the herpesvirus family, are particularly successful at evading or subverting the host immune response and have been shown to hijack GPCRs to favor their latent and lytic infection. In KSHV, the viral mimicker of cellular GPCR (vGPCR) is encoded by ORF74 (Figure 2F) and displays constitutive activity correlated with oncogenesis *in vitro* and *in vivo* (Arvanitakis et al., 1997; Bais et al., 1998; Jham and Montaner, 2010). In transgenic mice, vGPCR was shown to activate MAPKs, phospholipase C (PLC), PI3K and Akt (Smit et al., 2002), as well

as NF- κ B via the PI3K and Akt pathway (Pati et al., 2001), and subsequently release IL-6 and stimulate VEGF production in both paracrine and autocrine manners for cell growth and survival (Pati et al., 2001).

EBV also encodes a single viral GPCR, called BILF1, in the lytic replication phase (Figure 2E). Similar to vGPCR from KSHV, BILF1 is also able to transform cells *in vitro* and leads to tumor formation *in vivo* (Lyngaa et al., 2010), which may be dependent on a pattern of constitutive signaling through the G α I-activated pathway that in turn contributes to viral lytic replication (Paulsen et al., 2005). In the context of immune evasion, it has been shown that BILF1 can constitutively inhibit the phosphorylation of protein kinase R (PKR) (Beisser et al., 2005), and can activate transcriptional factors of NF- κ B and NF-AT to prevent the host cell antiviral response (Spiess et al., 2015). In addition, BILF1 reduces the levels of major histocompatibility complex (MHC) class I at the cell surface through the lysosomal degradation pathway (Zuo et al., 2009). Further studies have revealed that this function of BILF1 is dependent on the EKT signal motif located in its cytoplasmic tail (Zuo et al., 2011; Griffin et al., 2013). Similar to other viral GPCRs, BILF1 is able to hetero-oligomerize with other chemokine receptors like CXCR4, although BILF1 was previously demonstrated to be an orphan GPCR (Nijmeijer et al., 2010). These findings indicate that both GPCR homologs, encoded by EBV and KSHV, have highly similar functions in redirecting cellular GPCR signaling to favor for viral lytic replication.

VIRAL CYTOKINES AND CHEMOKINES

In addition to encoding their own viral membrane-associated proteins, it has been revealed that the genomes of EBV and KSHV also encode one or more viral cytokines or chemokines (Figure 1), allowing them to fully take over host membrane protein signaling. These viral genes have distinct homology to their cellular counterparts and could modulate cell signaling pathways to promote survival of the infected cell and escape from host antiviral immune surveillance. For example, EBV encodes a viral IL-10 (vIL-10) cytokine that is homologous to human IL-10 (Figure 3, left panel) and is expressed during primary or reactivated EBV infection phases for induction of B-cell transformation and growth (Miyazaki et al., 1993; Stuart et al., 1995; Bejarano and Masucci, 1998). To further prevent activation and recognition of T cells, vIL-10 has been found to downregulate the expression of the transporter protein TAP1, which associates with MHC class II antigens presented on monocytes, macrophages and B cells with EBV infection (de Waal Malefyt et al., 1991; Zeidler et al., 1997; Jochum et al., 2012), and also block interferon (IFN)- γ synthesis for

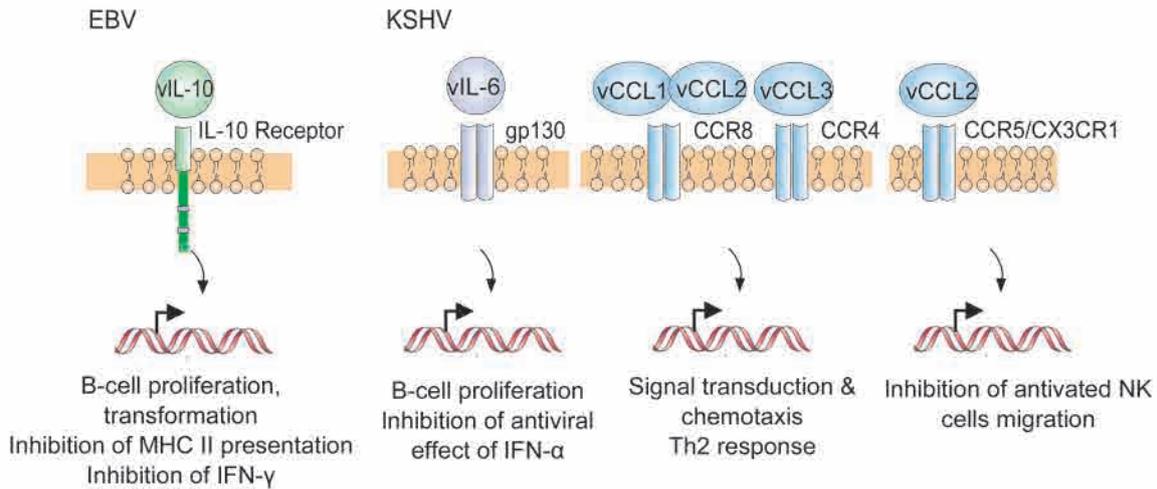


Figure 3. Schematic representation of the cytokines/chemokines encoded by EBV and KSHV and their related receptors and downstream signaling events.

enhancing EBV-infected B cell survival (Swaminathan et al., 1993; Suzuki et al., 1995). Interestingly, it has been shown that vIL-10 is highly conserved in various EBV isolates from gastric carcinomas and NPCs (Kanai et al., 2007; Chao et al., 2011) and may contribute to acute infection of permissive host cells by facilitating survival and dissemination of EBV-infected cells as seen in a murine herpesvirus model (Lindquester et al., 2014).

Although KSHV does not encode an ORF similar to the EBV vIL-10, a viral cytokine named vIL-6 is encoded. It has been demonstrated that vIL-6 is secreted from B cells after KSHV infection (Moore et al., 1996; Neipel et al., 1997; Nicholas et al., 1997) and can induce cellular IL-6 expression to promote cell proliferation of PEL cells (Mori et al., 2000) and block the antiviral effects of IFN- α (Chatterjee et al., 2002). However, distinct from cellular IL-6, vIL-6 only requires the gp130 subunit instead of both gp130 and IL-6 α receptors for signal transduction (Figure 3, middle panel) (Molden et al., 1997). Unexpectedly, in the context of KSHV-associated diseases, vIL-6 is highly expressed in PEL and MCD cells (Parravicini et al., 1997; Jones et al., 1999), but rarely in KS cells (Staskus et al., 1999; Parravicini et al., 2000).

Since chemokines play a profound role in leukocyte trafficking and development of adaptive immune responses, in addition to cytokines, KSHV also encodes several viral gene products with homology to cellular chemokines, known as vCCL1, vCCL2 and vCCL3 (Figure 3, right panels) (Boshoff et al., 1997). Unlike their cellular counterparts, vCCL1 and vCCL2 bind efficiently to the CCR8 receptor (Sozzani et al., 1998), while vCCL3 activates the CCR4 receptor for signal transduction and chemotaxis in monocytes (Stine et al., 2000). It

has been also proposed that vCCL2 and vCCL3 may play an immunomodulatory role in directing inflammation from the Th1 to Th2 type response for KSHV immune evasion (Sozzani et al., 1998; Singh et al., 2004). Recent studies further showed that vCCL2 is the only one of the four cytokine/chemokines encoded by KSHV that binds to natural killer (NK) cells through two different receptors, CX3CR1 and CCR5, which inhibits the migration of activated NK cells (Yamin et al., 2013). In contrast to KSHV, no viral counterpart of cellular chemokines in the EBV genome has been revealed so far. This indicates that there is a significant difference between EBV and KSHV in manipulating host cytokine and chemokine signals, and could explain why EBV and KSHV have different cell tropisms for infection, albeit both of them infect B cells.

REMARKS AND PERSPECTIVES

Recent progress in the tumor virology of both EBV and KSHV has given rise to important concepts relating to changes in the composition and function of host cell membrane proteins. It has been demonstrated that both EBV and KSHV have evolved a diverse array of viral genes to manipulate the host cell membrane, which encode unique viral proteins, viral homologues of cellular proteins and proteins involved in cell membrane signaling. The strategies utilized by these two viruses lead to deregulation of normal cellular pathways including apoptosis, antiviral immune surveillance and arrest in cell growth. As both EBV and KSHV have tropisms for different cell types, a certain subset of viral proteins located in the cell membrane might function in a cooperative manner to aid virus survival in different cellular en-

vironments, to ensure a life-long persistent infection within the host cells. In view of the fact that individual proteins of EBV or KSHV may or may not appear concurrently, the strategies that are used to subvert the host cell antiviral immune response and override cell-cycle checkpoints for cell growth or survival are often concordant. For instance, LMP1, encoded by EBV during latency, has a similar effect on cell growth as K1 encoded by KSHV during lytic replication; LMP2A has a similar function as K15 on inhibition of BCR activation; and the viral cytokines vIL10 and vIL6 encoded individually by EBV and KSHV promote B-cell proliferation. These phenomena support the notion that maintenance of viral latency with intermittent periods of productive viral replication is very common. Thus, the manipulation of the host cell membrane by viral proteins probably contributes to the progression and development of neoplastic diseases associated with EBV or KSHV infection.

In this review, we have discussed the composition and functions of host cell membrane proteins including homologs of receptors and cytokines. These viral proteins elicit many of the same phenotype as their cellular counterparts and therefore provide biological function in virus-infected cell types that do not normally express the corresponding cellular proteins. Our current knowledge about the hijacking of host cell membrane proteins and signaling by human γ -herpesviruses illustrates interesting similarities between proteins with apparently divergent functions at different stages of the viral life cycle. Despite similarities in the signaling pathways engaged by both EBV and KSHV, it is still very hard to answer why the same signaling pathway may result in transformation when targeted by some viral proteins but not others. Expression of membrane receptors, cytokines and chemokines during latency and lytic replication appears to be associated with an essential or at least contributory role in viral tumorigenesis, probably through mimicking physiological signals required for the survival of immune B or T cells.

The most interesting property that these host membrane proteins share is their ability to self-oligomerize for constitutive activation. However, it remains largely unclear whether oligomerization of these membrane proteins is caused by endogenous or exogenous ligands. Although it has been shown that no matter self- or ligand-induced oligomerization and interaction with host cellular factors, these viral transform membrane proteins could be adopted and modified cellular pathways for transforming host cells. In addition, both EBV and KSHV have also been demonstrated to manipulate the host cell membrane through deregulating the expression of both chemokines and cytokines or their receptors. For instance, EBV can induce expression of IL-8 and MIP-1 in human neutrophils (Mccoll et al., 1997); KSHV utilizes microRNA to reduce the production of inflamma-

tory-response cytokines (Abend et al., 2012) and dynamically regulate production of IL-6, TNF- α and MIP1 α in dendritic cells after KSHV primary infection (Hensler et al., 2009). This strategy is utilized to interact with other molecules that could directly impair the activation of host cell membrane receptors for viral pathogenesis. In conclusion, the functional elucidation of these cell membrane-associated proteins will provide potentially efficient strategies against EBV or KSHV-associated diseases.

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COMPLIANCE WITH ETHICS GUIDELINES

The authors declare that they have no conflict of interest. This article does not contain any studies with human or animal subjects performed by any of the authors.

REFERENCES

- Abend JR, Ramalingam D, Kieffer-Kwon P, Uldrick TS, Yarchoan R, Ziegelbauer JM. 2012. Kaposi's sarcoma-associated herpesvirus microRNAs target IRAK1 and MYD88, two components of the toll-like receptor/interleukin-1R signaling cascade, to reduce inflammatory-cytokine expression. *J Virol*, 86: 11663–11674.
- Amort M, Nachbauer B, Tuzlak S, Kieser A, Schepers A, Villunger A, Polacek N. 2015. Expression of the vault RNA protects cells from undergoing apoptosis. *Nat Commun*, 6: 7030.
- Anderson LJ, Longnecker R. 2008. EBV LMP2A provides a surrogate pre-B cell receptor signal through constitutive activation of the ERK/MAPK pathway. *J Gen Virol*, 89: 1563–1568.
- Arvanitakis L, Geras-Raaka E, Varma A, Gershengorn MC, Cesarman E. 1997. Human herpesvirus KSHV encodes a constitutively active G-protein-coupled receptor linked to cell proliferation. *Nature*, 385: 347–350.
- Bais C, Santomaso B, Coso O, Arvanitakis L, Raaka EG, Gutkind JS, Asch AS, Cesarman E, Gershengorn MC, Mesri EA. 1998. G-protein-coupled receptor of Kaposi's sarcoma-associated herpesvirus is a viral oncogene and angiogenesis activator. *Nature*, 391: 86–89.
- Bala K, Bosco R, Gramolelli S, Haas DA, Kati S, Pietrek M, Havemeier A, Yakushko Y, Singh VV, Dittrich-Breiholz O, Kracht M, Schulz TF. 2012. Kaposi's sarcoma herpesvirus K15

- protein contributes to virus-induced angiogenesis by recruiting PLCgamma1 and activating NFAT1-dependent RCAN1 expression. *PLoS Pathog*, 8: e1002927.
- Beisser PS, Verzijl D, Gruijthuijsen YK, Beuken E, Smit MJ, Leurs R, Bruggeman CA, Vink C. 2005. The Epstein-Barr virus BILF1 gene encodes a G protein-coupled receptor that inhibits phosphorylation of RNA-dependent protein kinase. *J Virol*, 79: 441–449.
- Bejarano MT, Masucci MG. 1998. Interleukin-10 abrogates the inhibition of Epstein-Barr virus-induced B-cell transformation by memory T-cell responses. *Blood*, 92: 4256–4262.
- Boshoff C, Endo Y, Collins PD, Takeuchi Y, Reeves JD, Schweickart VL, Siani MA, Sasaki T, Williams TJ, Gray PW, Moore PS, Chang Y, Weiss RA. 1997. Angiogenic and HIV-inhibitory functions of KSHV-encoded chemokines. *Science*, 278: 290–294.
- Bowser BS, DeWire SM, Damania B. 2002. Transcriptional regulation of the K1 gene product of Kaposi's sarcoma-associated herpesvirus. *J Virol*, 76: 12574–12583.
- Bowser BS, Morris S, Song MJ, Sun R, Damania B. 2006. Characterization of Kaposi's sarcoma-associated herpesvirus (KSHV) K1 promoter activation by Rta. *Virology*, 348: 309–327.
- Brinkmann MM, Glenn M, Rainbow L, Kieser A, Henke-Gendo C, Schulz TF. 2003. Activation of mitogen-activated protein kinase and NF-kappaB pathways by a Kaposi's sarcoma-associated herpesvirus K15 membrane protein. *J Virol*, 77: 9346–9358.
- Brinkmann MM, Pietrek M, Dittrich-Breiholz O, Kracht M, Schulz TF. 2007. Modulation of host gene expression by the K15 protein of Kaposi's sarcoma-associated herpesvirus. *J Virol*, 81: 42–58.
- Burkhardt AL, Bolen JB, Kieff E, Longnecker R. 1992. An Epstein-Barr virus transformation-associated membrane protein interacts with src family tyrosine kinases. *J Virol*, 66: 5161–5167.
- Calderwood MA, Venkatesan K, Xing L, Chase MR, Vazquez A, Holthaus AM, Ewence AE, Li N, Hirozane-Kishikawa T, Hill DE. 2007. Epstein-Barr virus and virus human protein interaction maps. *Proc Natl Acad Sci U S A*, 104: 7606–7611.
- Caldwell RG, Wilson JB, Anderson SJ, Longnecker R. 1998. Epstein-Barr virus LMP2A drives B cell development and survival in the absence of normal B cell receptor signals. *Immunity*, 9: 405–411.
- Cen O, Longnecker R. 2015. Latent Membrane Protein 2 (LMP2). *Curr Top Microbiol Immunol*, 391: 151–180.
- Cesarman E, Chang Y, Moore PS, Said JW, Knowles DM. 1995. Kaposi's sarcoma-associated herpesvirus-like DNA sequences in AIDS-related body-cavity-based lymphomas. *N Engl J Med*, 332: 1186–1191.
- Chakraborty S, Veettil MV, Chandran B. 2012. Kaposi's sarcoma associated herpesvirus entry into target cells. *Front Microbiol*, 3: 6.
- Chang Y, Cesarman E, Pessin MS, Lee F, Culpepper J, Knowles DM, Moore PS. 1994. Identification of herpesvirus-like DNA sequences in AIDS-associated Kaposi's sarcoma. *Science*, 266: 1865–1869.
- Chao Y, Jing Y, Jia Y, Wang Y, Zhao C, Luo B. 2011. Conservation and mutation of viral interleukin-10 gene in gastric carcinomas and nasopharyngeal carcinomas. *J Med Virol*, 83: 644–650.
- Chatterjee M, Osborne J, Bestetti G, Chang Y, Moore PS. 2002. Viral IL-6-induced cell proliferation and immune evasion of interferon activity. *Science*, 298: 1432–1435.
- Chazal N, Gerlier D. 2003. Virus entry, assembly, budding, and membrane rafts. *Microbiol Mol Biol Rev*, 67: 226–237.
- de Waal Malefyt R, Haanen J, Spits H, Roncarolo MG, te Velde A, Figdor C, Johnson K, Kastelein R, Yssel H, de Vries JE. 1991. Interleukin 10 (IL-10) and viral IL-10 strongly reduce antigen-specific human T cell proliferation by diminishing the antigen-presenting capacity of monocytes via downregulation of class II major histocompatibility complex expression. *J Exp Med*, 174: 915–924.
- Epstein MA, Achong BG, Barr YM. 1964. Virus Particles in Cultured Lymphoblasts from Burkitt's Lymphoma. *Lancet*, 1: 702–703.
- Fish K, Chen J, Longnecker R. 2014. Epstein-Barr virus latent membrane protein 2A enhances MYC-driven cell cycle progression in a mouse model of B lymphoma. *Blood*, 123: 530–540.
- Fotheringham JA, Coalson NE, Raab-Traub N. 2012. Epstein-Barr virus latent membrane protein-2A induces ITAM/Syk- and Akt-dependent epithelial migration through alphav-integrin membrane translocation. *J Virol*, 86: 10308–10320.
- Fox CP, Haigh TA, Taylor GS, Long HM, Lee SP, Shannon-Lowe C, O'Connor S, Bollard CM, Iqbal J, Chan WC, Rickinson AB, Bell AI, Rowe M. 2010. A novel latent membrane 2 transcript expressed in Epstein-Barr virus-positive NK- and T-cell lymphoproliferative disease encodes a target for cellular immunotherapy. *Blood*, 116: 3695–3704.
- Fruehling S, Longnecker R. 1997. The immunoreceptor tyrosine-based activation motif of Epstein-Barr virus LMP2A is essential for blocking BCR-mediated signal transduction. *Virology*, 235: 241–251.
- Fukuda M, Kawaguchi Y. 2014. Role of the immunoreceptor tyrosine-based activation motif of latent membrane protein 2A (LMP2A) in Epstein-Barr virus LMP2A-induced cell transformation. *J Virol*, 88: 5189–5194.
- Fukuda M, Longnecker R. 2005. Epstein-Barr virus (EBV) latent membrane protein 2A regulates B-cell receptor-induced apoptosis and EBV reactivation through tyrosine phosphorylation. *J Virol*, 79: 8655–8660.
- Glenn M, Rainbow L, Aurade F, Davison A, Schulz TF. 1999. Identification of a spliced gene from Kaposi's sarcoma-associated herpesvirus encoding a protein with similarities to latent membrane proteins 1 and 2A of Epstein-Barr virus. *J Virol*, 73: 6953–6963.
- Goldberg DM, Riordan JR. 1986. Role of membranes in disease. *Clin Physiol Biochem*, 4: 305–336.
- Gramolelli S, Weidner-Glunde M, Abere B, Viejo-Borbolla A, Bala K, Ruckert J, Kremmer E, Schulz TF. 2015. Inhibiting the Recruitment of PLCgamma1 to Kaposi's Sarcoma Herpesvirus K15 Protein Reduces the Invasiveness and Angiogenesis of Infected Endothelial Cells. *PLoS Pathog*, 11: e1005105.
- Griffin BD, Gram AM, Mulder A, Van Leeuwen D, Claas FH, Wang F, Rensing ME, Wiertz E. 2013. EBV BILF1 evolved to downregulate cell surface display of a wide range of HLA class I molecules through their cytoplasmic tail. *J Immunol*, 190: 1672–1684.
- Gujer C, Chatterjee B, Landtwing V, Raykova A, McHugh D, Munz C. 2015. Animal models of Epstein Barr virus infection. *Curr Opin Virol*, 13: 6–10.
- Han J, Chen JN, Zhang ZG, Li HG, Ding YG, Du H, Shao CK. 2012. Sequence variations of latent membrane protein 2A in Epstein-Barr virus-associated gastric carcinomas from Guang-

- zhou, southern China. *PLoS One*, 7: e34276.
- Hatzivassiliou E, Miller WE, Raab-Traub N, Kieff E, Mosialos G. 1998. A fusion of the EBV latent membrane protein-1 (LMP1) transmembrane domains to the CD40 cytoplasmic domain is similar to LMP1 in constitutive activation of epidermal growth factor receptor expression, nuclear factor-kappa B, and stress-activated protein kinase. *J Immunol*, 160: 1116–1121.
- Heaton NS, Randall G. 2011. Multifaceted roles for lipids in viral infection. *Trends Microbiol*, 19: 368–375.
- Hensler HR, Rappocciolo G, Rinaldo CR, Jenkins FJ. 2009. Cytokine production by human herpesvirus 8-infected dendritic cells. *J Gen Virol*, 90: 79–83.
- Izumi KM, Kaye KM, Kieff ED. 1997. The Epstein-Barr virus LMP1 amino acid sequence that engages tumor necrosis factor receptor associated factors is critical for primary B lymphocyte growth transformation. *Proc Natl Acad Sci U S A*, 94: 1447–1452.
- Jham BC, Montaner S. 2010. The Kaposi's sarcoma-associated herpesvirus G protein-coupled receptor: Lessons on dysregulated angiogenesis from a viral oncogene. *J Cell Biochem*, 110: 1–9.
- Jochum S, Moosmann A, Lang S, Hammerschmidt W, Zeidler R. 2012. The EBV immunoevasins vIL-10 and BNLF2a protect newly infected B cells from immune recognition and elimination. *PLoS Pathog*, 8: e1002704.
- Jones KD, Aoki Y, Chang Y, Moore PS, Yarchoan R, Tosato G. 1999. Involvement of interleukin-10 (IL-10) and viral IL-6 in the spontaneous growth of Kaposi's sarcoma herpesvirus-associated infected primary effusion lymphoma cells. *Blood*, 94: 2871–2879.
- Kanai K, Satoh Y, Yamanaka H, Kawaguchi A, Horie K, Sugata K, Hoshikawa Y, Sata T, Sairenji T. 2007. The vIL-10 gene of the Epstein-Barr virus (EBV) is conserved in a stable manner except for a few point mutations in various EBV isolates. *Virus Genes*, 35: 563–569.
- Lagunoff M, Ganem D. 1997. The structure and coding organization of the genomic termini of Kaposi's sarcoma-associated herpesvirus. *Virology*, 236: 147–154.
- Lagunoff M, Majeti R, Weiss A, Ganem D. 1999. Deregulated signal transduction by the K1 gene product of Kaposi's sarcoma-associated herpesvirus. *Proc Natl Acad Sci U S A*, 96: 5704–5709.
- Lee H, Guo J, Li M, Choi JK, DeMaria M, Rosenzweig M, Jung JU. 1998a. Identification of an immunoreceptor tyrosine-based activation motif of K1 transforming protein of Kaposi's sarcoma-associated herpesvirus. *Mol Cell Biol*, 18: 5219–5228.
- Lee H, Veazey R, Williams K, Li M, Guo J, Neipel F, Fleckenstein B, Lackner A, Desrosiers RC, Jung JU. 1998b. Deregulation of cell growth by the K1 gene of Kaposi's sarcoma-associated herpesvirus. *Nat Med*, 4: 435–440.
- Lindquister GJ, Greer KA, Stewart JP, Sample JT. 2014. Epstein-Barr virus IL-10 gene expression by a recombinant murine gammaherpesvirus in vivo enhances acute pathogenicity but does not affect latency or reactivation. *Herpesviridae*, 5: 1.
- Lu J, Lin WH, Chen SY, Longnecker R, Tsai SC, Chen CL, Tsai CH. 2006. Syk tyrosine kinase mediates Epstein-Barr virus latent membrane protein 2A-induced cell migration in epithelial cells. *J Biol Chem*, 281: 8806–8814.
- Lyngaa R, Norregaard K, Kristensen M, Kubale V, Rosenkilde MM, Kledal TN. 2010. Cell transformation mediated by the Epstein-Barr virus G protein-coupled receptor BILF1 is dependent on constitutive signaling. *Oncogene*, 29: 4388–4398.
- Ma SD, Xu X, Plowshay J, Ranheim EA, Burlingham WJ, Jensen JL, Asimakopoulos F, Tang W, Gulley ML, Cesarman E, Gumperz JE, Kenney SC. 2015. LMP1-deficient Epstein-Barr virus mutant requires T cells for lymphomagenesis. *J Clin Invest*, 125: 304–315.
- Mazzoni M, Mercer J. 2014. Lipid interactions during virus entry and infection. *Cell Microbiol*, 16: 1493–1502.
- McCull SR, Roberge CJ, Laroche B, Gosselin J. 1997. EBV induces the production and release of IL-8 and macrophage inflammatory protein-1 alpha in human neutrophils. *J Immunol*, 159: 6164–6168.
- Meckes DG, Jr., Shair KH, Marquitz AR, Kung CP, Edwards RH, Raab-Traub N. 2010. Human tumor virus utilizes exosomes for intercellular communication. *Proc Natl Acad Sci U S A*, 107: 20370–20375.
- Merchant M, Caldwell RG, Longnecker R. 2000. The LMP2A ITAM is essential for providing B cells with development and survival signals in vivo. *J Virol*, 74: 9115–9124.
- Mesri EA, Cesarman E, Boshoff C. 2010. Kaposi's sarcoma and its associated herpesvirus. *Nat Rev Cancer*, 10: 707–719.
- Miller CL, Burkhardt AL, Lee JH, Stealey B, Longnecker R, Bolen JB, Kieff E. 1995. Integral membrane protein 2 of Epstein-Barr virus regulates reactivation from latency through dominant negative effects on protein-tyrosine kinases. *Immunity*, 2: 155–166.
- Miller CL, Lee JH, Kieff E, Longnecker R. 1994. An integral membrane protein (LMP2) blocks reactivation of Epstein-Barr virus from latency following surface immunoglobulin crosslinking. *Proc Natl Acad Sci U S A*, 91: 772–776.
- Miller CL, Longnecker R, Kieff E. 1993. Epstein-Barr virus latent membrane protein 2A blocks calcium mobilization in B lymphocytes. *J Virol*, 67: 3087–3094.
- Miyazaki I, Cheung RK, Dosch HM. 1993. Viral interleukin 10 is critical for the induction of B cell growth transformation by Epstein-Barr virus. *J Exp Med*, 178: 439–447.
- Molden J, Chang Y, You Y, Moore PS, Goldsmith MA. 1997. A Kaposi's sarcoma-associated herpesvirus-encoded cytokine homolog (vIL-6) activates signaling through the shared gp130 receptor subunit. *J Biol Chem*, 272: 19625–19631.
- Moore PS, Boshoff C, Weiss RA, Chang Y. 1996. Molecular mimicry of human cytokine and cytokine response pathway genes by KSHV. *Science*, 274: 1739–1744.
- Mori Y, Nishimoto N, Ohno M, Inagi R, Dhepakson P, Amou K, Yoshizaki K, Yamanishi K. 2000. Human herpesvirus 8-encoded interleukin-6 homologue (viral IL-6) induces endogenous human IL-6 secretion. *J Med Virol*, 61: 332–335.
- Motsch N, Pfuhl T, Mrazek J, Barth S, Grasser FA. 2007. Epstein-Barr virus-encoded latent membrane protein 1 (LMP1) induces the expression of the cellular microRNA miR-146a. *RNA Biol*, 4: 131–137.
- Neipel F, Albrecht JC, Ensser A, Huang YQ, Li JJ, Friedman-Kien AE, Fleckenstein B. 1997. Human herpesvirus 8 encodes a homolog of interleukin-6. *J Virol*, 71: 839–842.
- Nicholas J, Ruvolo VR, Burns WH, Sandford G, Wan X, Ciuffo D, Hendrickson SB, Guo HG, Hayward GS, Reitz MS. 1997. Kaposi's sarcoma-associated human herpesvirus-8 encodes homologues of macrophage inflammatory protein-1 and interleukin-6. *Nat Med*, 3: 287–292.
- Nijmeijer S, Leurs R, Smit MJ, Vischer HF. 2010. The Epstein-Barr virus-encoded G protein-coupled receptor BILF1 heterooligomerizes with human CXCR4, scavenges Gα proteins, and constitutively impairs CXCR4 functioning. *J Biol Chem*,

- 285: 29632–29641.
- Parravicini C, Chandran B, Corbellino M, Berti E, Paulli M, Moore PS, Chang Y. 2000. Differential viral protein expression in Kaposi's sarcoma-associated herpesvirus-infected diseases: Kaposi's sarcoma, primary effusion lymphoma, and multicentric Castlemans disease. *Am J Pathol*, 156: 743–749.
- Parravicini C, Corbellino M, Paulli M, Magrini U, Lazzarino M, Moore PS, Chang Y. 1997. Expression of a virus-derived cytokine, KSHV vIL-6, in HIV-seronegative Castlemans disease. *Am J Pathol*, 151: 1517–1522.
- Pati S, Cavrois M, Guo HG, Foulke JS, Jr., Kim J, Feldman RA, Reitz M. 2001. Activation of NF- κ B by the human herpesvirus 8 chemokine receptor ORF74: evidence for a paracrine model of Kaposi's sarcoma pathogenesis. *J Virol*, 75: 8660–8673.
- Paulsen SJ, Rosenkilde MM, Eugen-Olsen J, Kledal TN. 2005. Epstein-Barr virus-encoded BILF1 is a constitutively active G protein-coupled receptor. *J Virol*, 79: 536–546.
- Pietrek M, Brinkmann MM, Glowacka I, Enlund A, Havemeier A, Dittrich-Breiholz O, Kracht M, Lewitzky M, Saksela K, Feller SM, Schulz TF. 2010. Role of the Kaposi's sarcoma-associated herpesvirus K15 SH3 binding site in inflammatory signaling and B-cell activation. *J Virol*, 84: 8231–8240.
- Portis T, Longnecker R. 2004. Epstein-Barr virus (EBV) LMP2A mediates B-lymphocyte survival through constitutive activation of the Ras/PI3K/Akt pathway. *Oncogene*, 23: 8619–8628.
- Qu L, Green M, Webber S, Reyes J, Ellis D, Rowe D. 2000. Epstein-Barr virus gene expression in the peripheral blood of transplant recipients with persistent circulating virus loads. *J Infect Dis*, 182: 1013–1021.
- Riethmüller J, Riehle A, Grassmé H, Gulbins E. 2006. Membrane rafts in host–pathogen interactions. *Biochim Biophys Acta*, 1758: 2139–2147.
- Rosenbaum DM, Rasmussen SG, Kobilka BK. 2009. The structure and function of G-protein-coupled receptors. *Nature*, 459: 356–363.
- Singh UP, Singh S, Ravichandran P, Taub DD, Lillard JW, Jr. 2004. Viral macrophage-inflammatory protein-II: a viral chemokine that differentially affects adaptive mucosal immunity compared with its mammalian counterparts. *J Immunol*, 173: 5509–5516.
- Siouda M, Frecha C, Accardi R, Yue J, Cuenin C, Gruffat H, Manet E, Herceg Z, Sylla BS, Tommasino M. 2014. Epstein-Barr virus down-regulates tumor suppressor DOK1 expression. *PLoS Pathog*, 10: e1004125.
- Smit MJ, Verzijl D, Casarosa P, Navis M, Timmerman H, Leurs R. 2002. Kaposi's sarcoma-associated herpesvirus-encoded G protein-coupled receptor ORF74 constitutively activates p44/p42 MAPK and Akt via G(i) and phospholipase C-dependent signaling pathways. *J Virol*, 76: 1744–1752.
- Sozzani S, Luini W, Bianchi G, Allavena P, Wells TN, Napolitano M, Bernardini G, Vecchi A, D'Ambrosio D, Mazzeo D, Sinigaglia F, Santoni A, Maggi E, Romagnani S, Mantovani A. 1998. The viral chemokine macrophage inflammatory protein-II is a selective Th2 chemoattractant. *Blood*, 92: 4036–4039.
- Spear PG, Longnecker R. 2003. Herpesvirus entry: an update. *J Virol*, 77: 10179–10185.
- Spieß K, Fares S, Sparre-Ulrich AH, Hilgenberg E, Jarvis MA, Ehlers B, Rosenkilde MM. 2015. Identification and functional comparison of seven-transmembrane G-protein-coupled BILF1 receptors in recently discovered nonhuman primate lymphocryptoviruses. *J Virol*, 89: 2253–2267.
- Staskus KA, Sun R, Miller G, Racz P, Jaslowski A, Metroka C, Brett-Smith H, Haase AT. 1999. Cellular tropism and viral interleukin-6 expression distinguish human herpesvirus 8 involvement in Kaposi's sarcoma, primary effusion lymphoma, and multicentric Castlemans disease. *J Virol*, 73: 4181–4187.
- Stewart S, Dawson CW, Takada K, Curnow J, Moody CA, Sixbey JW, Young LS. 2004. Epstein-Barr virus-encoded LMP2A regulates viral and cellular gene expression by modulation of the NF- κ B transcription factor pathway. *Proc Natl Acad Sci U S A*, 101: 15730–15735.
- Stine JT, Wood C, Hill M, Epp A, Raport CJ, Schweickart VL, Endo Y, Sasaki T, Simmons G, Boshoff C, Clapham P, Chang Y, Moore P, Gray PW, Chantry D. 2000. KSHV-encoded CC chemokine vMIP-1 is a CCR4 agonist, stimulates angiogenesis, and selectively chemoattracts TH2 cells. *Blood*, 95: 1151–1157.
- Stuart AD, Stewart JP, Arrand JR, Mackett M. 1995. The Epstein-Barr virus encoded cytokine viral interleukin-10 enhances transformation of human B lymphocytes. *Oncogene*, 11: 1711–1719.
- Suzuki T, Tahara H, Narula S, Moore KW, Robbins PD, Lotze MT. 1995. Viral interleukin 10 (IL-10), the human herpes virus 4 cellular IL-10 homologue, induces local anergy to allogeneic and syngeneic tumors. *J Exp Med*, 182: 477–486.
- Swaminathan S, Hesselton R, Sullivan J, Kieff E. 1993. Epstein-Barr virus recombinants with specifically mutated BCRF1 genes. *J Virol*, 67: 7406–7413.
- Thorley-Lawson DA, Hawkins JB, Tracy SI, Shapiro M. 2013. The pathogenesis of Epstein-Barr virus persistent infection. *Curr Opin Virol*, 3: 227–232.
- Tomlinson CC, Damania B. 2004. The K1 protein of Kaposi's sarcoma-associated herpesvirus activates the Akt signaling pathway. *J Virol*, 78: 1918–1927.
- Tomlinson CC, Damania B. 2008. Critical role for endocytosis in the regulation of signaling by the Kaposi's sarcoma-associated herpesvirus K1 protein. *J Virol*, 82: 6514–6523.
- Veetil MV, Bandyopadhyay C, Dutta D, Chandran B. 2014. Interaction of KSHV with host cell surface receptors and cell entry. *Viruses*, 6: 4024–4046.
- Wang D, Liebowitz D, Kieff E. 1985. An EBV membrane protein expressed in immortalized lymphocytes transforms established rodent cells. *Cell*, 43: 831–840.
- Wang D, Liebowitz D, Wang F, Gregory C, Rickinson A, Larson R, Springer T, Kieff E. 1988. Epstein-Barr virus latent infection membrane protein alters the human B-lymphocyte phenotype: deletion of the amino terminus abolishes activity. *J Virol*, 62: 4173–4184.
- Wang L, Brinkmann MM, Pietrek M, Ottinger M, Dittrich-Breiholz O, Kracht M, Schulz TF. 2007. Functional characterization of the M-type K15-encoded membrane protein of Kaposi's sarcoma-associated herpesvirus. *J Gen Virol*, 88: 1698–1707.
- Wang L, Dittmer DP, Tomlinson CC, Fakhari FD, Damania B. 2006. Immortalization of primary endothelial cells by the K1 protein of Kaposi's sarcoma-associated herpesvirus. *Cancer Res*, 66: 3658–3666.
- Wang L, Wakisaka N, Tomlinson CC, DeWire SM, Krall S, Pagano JS, Damania B. 2004. The Kaposi's sarcoma-associated herpesvirus (KSHV/HHV-8) K1 protein induces expression of angiogenic and invasion factors. *Cancer Res*, 64: 2774–2781.
- Wen KW, Damania B. 2010. Hsp90 and Hsp40/Erdj3 are required for the expression and anti-apoptotic function of KSHV K1. On-

- cogene, 29: 3532–3544.
- Xiao L, Hu ZY, Dong X, Tan Z, Li W, Tang M, Chen L, Yang L, Tao Y, Jiang Y, Li J, Yi B, Li B, Fan S, You S, Deng X, Hu F, Feng L, Bode AM, Dong Z, Sun LQ, Cao Y. 2014. Targeting Epstein-Barr virus oncoprotein LMP1-mediated glycolysis sensitizes nasopharyngeal carcinoma to radiation therapy. *Oncogene*, 33: 4568–4578.
- Yamin R, Kaynan NS, Glasner A, Vitenshtein A, Tsukerman P, Bauman Y, Ophir Y, Elias S, Bar-On Y, Gur C, Mandelboim O. 2013. The viral KSHV chemokine vMIP-II inhibits the migration of Naïve and activated human NK cells by antagonizing two distinct chemokine receptors. *PLoS Pathog*, 9: e1003568.
- Zeidler R, Eissner G, Meissner P, Uebel S, Tampe R, Lazis S, Hammerschmidt W. 1997. Downregulation of TAP1 in B lymphocytes by cellular and Epstein-Barr virus-encoded interleukin-10. *Blood*, 90: 2390–2397.
- Zhang Z, Chen W, Sanders MK, Brulois KF, Dittmer DP, Damania B. 2016. The K1 protein of Kaposi's sarcoma-associated herpesvirus (KSHV) augments viral lytic replication. *J Virol*. pii: JVI.03102-15.
- Zong JC, Ciufu DM, Alcendor DJ, Wan X, Nicholas J, Browning PJ, Rady PL, Tying SK, Orenstein JM, Rabkin CS, Su IJ, Powell KF, Croxson M, Foreman KE, Nickoloff BJ, Alkan S, Hayward GS. 1999. High-level variability in the ORF-K1 membrane protein gene at the left end of the Kaposi's sarcoma-associated herpesvirus genome defines four major virus subtypes and multiple variants or clades in different human populations. *J Virol*, 73: 4156–4170.
- Zuo J, Currin A, Griffin BD, Shannon-Lowe C, Thomas WA, Rensing ME, Wiertz EJ, Rowe M. 2009. The Epstein-Barr virus G-protein-coupled receptor contributes to immune evasion by targeting MHC class I molecules for degradation. *PLoS Pathog*, 5: e1000255.
- Zuo J, Quinn LL, Tamblyn J, Thomas WA, Feederle R, Delecluse HJ, Hislop AD, Rowe M. 2011. The Epstein-Barr virus-encoded BILF1 protein modulates immune recognition of endogenously processed antigen by targeting major histocompatibility complex class I molecules trafficking on both the exocytic and endocytic pathways. *J Virol*, 85: 1604–1614.